

Fast detection of snakes and emotional faces in the macaque amygdala

(マカクサル扁桃体ニューロンはヘビおよび情動表情を素早く検出する)

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Abstract

Primate vision is reported to detect snakes and emotional faces faster than many other tested stimuli. Because the amygdala has been implicated in avoidance and emotional behaviors to biologically relevant stimuli and has neural connections with subcortical nuclei involved with vision, amygdalar neurons would be sensitive to snakes and emotional faces. In this study, neuronal activity in the amygdala was recorded from Japanese macaques (*Macaca fuscata*) during discrimination of eight categories of visual stimuli including snakes, monkey faces, human faces, carnivores, raptors, non-predators, monkey hands, and simple figures. Of 527 amygdalar neurons, 95 responded to one or more stimuli. Response characteristics of the amygdalar neurons indicated that they were more sensitive to the snakes and emotional faces than other stimuli. Response magnitudes and latencies of amygdalar neurons to snakes and monkey faces were stronger and faster than those to the other categories of stimuli, respectively. Furthermore, response magnitudes to the low pass-filtered snake images were larger than those to scrambled snake images. Finally, analyses of population activity of amygdalar neurons suggest that snakes and emotional faces were represented separately from the other stimuli during the 50-100 ms period from stimulus onset, and neutral faces during the 100-150 ms period. These response characteristics indicate that the amygdala processes fast and coarse visual information from emotional faces and snakes (but not other predators of primates) among the eight categories of the visual stimuli, and suggest that, like anthropoid primate visual systems, the amygdala has been shaped over evolutionary time to detect appearance of potentially threatening stimuli including both emotional faces and snakes, the first of the modern predators of primates.

Keywords: macaque, amygdala, single-unit activity, snakes, emotional faces

1. Introduction

Rapid defensive responses are often required to avoid predators. Neural systems for such behaviors have been argued to be controlled by an evolved subcortical “fear module” that in mammals includes the superior colliculus (SC) and pulvinar, two nuclei involved with vision, and the amygdala (Öhman and Mineka, 2001; Isbell 2006, 2009). Both the SC and pulvinar receive visual inputs from the retinal ganglion cells, while the pulvinar receives visual inputs from the superficial and deep layers of the SC (Soares et al., 2017; Isa et al., 2021). These retinal ganglion cells projecting to the SC mainly convey magnocellular (M) and kinocellular (K) channels of visual information with low spatial resolution (Waleszczyk et al., 2004; Grünert et al., 2021). The pulvinar further projects to at least lateral part of the amygdala in macaques (Elorette et al., 2018). This SC-pulvinar-amygdalar pathway appears to unconsciously detect appearance/movement of potentially threatening stimuli including phylogenetically salient stimuli in shorter latencies than the cortical visual system (Tamietto and de Gelder, 2010; Soares et al., 2017; McFadyen et al., 2020). Anthropoid primates (including humans) are highly dependent on vision for detecting danger. It has been argued that, as the first modern predator of primates, snakes have long exerted strong evolutionary pressure on primate visual systems to increase sensitivity to visual cues provided by snakes (Isbell, 2006, 2009). Growing evidence consistently supports this. For example, naïve monkeys and humans detect snakes more quickly than other objects under a variety of conditions (Nelson et al., 2003; Kawai and Koda, 2016; Bertels et al., 2020).

As highly social mammals, anthropoid primates must also respond quickly to conspecifics, especially when threatened. The subcortical “fear module” has been implicated in the ability of people with cortical lesions showing affective blindsight to unconsciously discriminate emotional faces (Morris et al., 2001; de Gelder et al., 2005; Penga et al., 2005; Tamietto et al., 2012). A neurophysiological study reported that monkey pulvinar neurons in macaque monkeys responded more quickly and more strongly to both of these phylogenetically salient stimuli than to neutral objects (monkey hands and geometric shapes), as expected of nuclei in the “fear module” (Le et al., 2013). It is likely that the pulvinar’s biased responses for images of snakes and emotional faces arose via the superior colliculus as the pulvinar receives visual inputs from the superior colliculus (Le et al., 2013). In rodents, the amygdala, the other core structure of the “fear module”, plays a crucial role in mediating defensive responses to unconditioned stimuli such as predator-relevant stimuli (Martinez et al., 2011; Root et al., 2014; Isosaka et al., 2015). However, the responsiveness of amygdalar neurons to phylogenetically salient stimuli in primates, especially snakes, is currently unknown.

In this study, I attempted to close this gap in knowledge by comparing neuronal responses to snakes and emotional faces as salient stimuli against other presumably less salient visual stimuli in the amygdala of the Japanese macaque (*Macaca fuscata*) as a model for anthropoid primates. To analyze amygdalar neuronal sensitivity to innate biological significance of visual stimuli, especially snakes, amygdalar neurons were tested with eight categories of the visual stimuli [snakes, monkey faces (emotional and neutral faces), human faces (emotional and neutral faces), monkey hands, non-predators, raptors, carnivores, and simple figures]. The main predators of primates are carnivores, raptors, and snakes, of which snakes are the first of the modern predators of primates (Isbell, 1994), suggesting that snakes might shape the visual system of primates for a longer time than the other predators. Facial stimuli including emotional faces were also included as visual stimuli to test amygdalar neurons, since these stimuli were reported to be biologically salient (Öhman et al., 2001; Hershler and Hochstein, 2005; Hahn et al., 2006; Langton et al., 2008). Non-predators and simple figures were used as control stimuli that are supposed to have less innate biological

significance. However, stimuli with physical salience such as onset/offset of intense light, to which the subcortical visual pathway are also sensitive (Itti and Koch, 2001; Yoshida et al., 2012), were not used in the present study. Amygdalar neurons were also tested with low- and high-pass filtered snake images as well as scrambled snake images. The subcortical pathway including the amygdala, which processes fast and coarse information, is reported to be more responsive to low-pass filtered stimuli (Vuilleumier et al. 2003; Le et al., 2013; Gomes et al., 2018). Here, I report that macaque amygdalar neurons responded faster and stronger to snakes than to other potential predators for primates, and that they responded faster and stronger to emotional faces compared to neutral faces. The results also indicated that amygdalar neurons were more sensitive to low spatial frequency components of snake images compared to high spatial frequency components of those stimuli, and scrambling of snake images decreased the responses. These results suggest that, like the pulvinar, the anthropoid primate amygdala has evolved to receive fast and coarse visual information about evolutionarily salient stimuli that have the potential to affect primate survival.

My doctoral dissertation based on my academic thesis has been published on an open journal (Dinh et al., 2022).

2. Materials and Methods

2.1. Subjects

Two adult male Japanese macaques (Monkey1 and Monkey2), weighing 7.3 and 8.6 kg (6 years old at the time of training and recording), were used in this experiment. Both monkeys were not used in the previous experiments including Le et al. (2013) and Dinh et al. (2018). They were born in a breeding facility in the Primate Research Institute in Kyoto University, Japan, and reared in a group cage (5-6 monkeys per cage: Monkey1), where one side of the cage was in contact with the external environment and monkeys could freely see outside, or in a colony in an outer enclosed area (Monkey2). They were transferred to University of Toyama at the age of three, and individually housed with free access to monkey chow and water. They were also given vegetables and fruits. Environmental enrichment (toys) was provided daily. They experienced no stressful procedures for the whole duration. The monkeys were deprived of water in their home cages during training and recording sessions and received supplemental water and vegetables after each day's session at the age of six. The monkeys were treated according to the Guidelines for the Care and Use of Laboratory Animals of the University of Toyama as well as the United States Public Health Service Policy on Human Care and Use of Laboratory Animals, the National Institutes of Health Guide for the Care and Use of Laboratory Animals. According to Japanese law, the Committee for Animal Experiments and Ethics at the University of Toyama reviewed and approved the experimental protocol and procedures in this study (permit no.: A2016MED-2, A2019MED-18, and A2022MED-1).

2.2. Visual stimuli

The same visual stimuli that were used in a previous study (Dinh et al., 2018) were used in this study except that a cat in the previous study was replaced with a cow in the present study. Figure 1Aa shows eight categories of the visual stimuli used in the present study: snakes, monkey faces (emotional and neutral faces), human faces (emotional and neutral faces), monkey hands, non-predators, raptors, carnivores, and simple figures.

The color stimuli were 8-bits digitized RGB color-scale images (227×227 pixels). The luminance of the color stimuli was very similar (6.005-6.445 cd/m^2). The luminance of the white areas inside the simple figures was 36.5 cd/m^2 . Michelson contrast was not significantly different among the eight categories of the visual stimuli (Dinh et al., 2018).

These stimuli were presented at the center of a display (resolution: 640×480 pixels) with a black background of 0.7 cd/m^2 . The size of the stimulus area was $5\text{-}7 \times 5\text{-}7^\circ$.

2.3. Transformed visual stimuli

The transformed visual stimuli were introduced to analyze the characteristics of the stimuli that amygdalar neurons responded to (Figure 1Ab). According to previous studies (Vuilleumier et al., 2003; Rotshtein et al., 2007), a low-pass filter (LPF: 6 cycles/image) and a high-pass filter (HPF: 20 cycles/image) were used to modify spatial frequencies of the stimuli. The original visual stimuli were processed in the same way as a previous study (Le et al., 2013) to obtain the low- and high-pass filtered images.

Following a previous study (Honey et al., 2008), the visual images were scrambled using two different methods (Fourier and Wavelet scrambling). In Fourier scrambling, the positions (i.e., phases) of all spatial frequency components were randomized while the Fourier 2-D amplitude spectrum across orientations and spatial frequencies were maintained. In wavelet scrambling, the orientations of local spatial frequency components were randomized while the local wavelet coefficient values were maintained. These transformed visual stimuli maintained almost completely the global low-level properties of the original images (i.e., global contrast, luminance, spatial frequency amplitude spectrum, color) (Honey et al., 2008). These images were transformed using MatLab.

Some visual stimuli were transformed (adjusted visual stimuli) using the SHINE toolbox on MatLab (Willenbockel et al., 2010; Railo et al., 2016) so that low-level properties (the color histogram and the amplitude spectra of each color channel of RGB) of the visual stimuli were equalized with means of the low-level properties of the snake stimuli. However, the equalization of the properties cannot be perfect because of limitations in the method (Willenbockel et al., 2010). Therefore, to determine if the equalization removed differences of low-level properties among the stimulus categories, the following low-level properties of the adjusted stimuli were statistically compared (Le et al., 2016). Michelson contrast was compared among the five categories by one-way ANOVA (the categories: the original snakes, adjusted raptors, adjusted human faces, adjusted monkey faces, and adjusted carnivores). Michelson contrast of each category of the adjusted stimuli was also directly compared with the original snake stimuli by unpaired *t*-test. A color histogram consisting of four equally spaced bins was calculated for each color channel. Each bin of the histograms was statistically compared with one-way ANOVA and unpaired *t*-test in the same way as described above. Power spectrum averaged overall orientation (Simoncelli and Olshausen, 2001) was calculated for each color channel. The total power of low and high spatial frequency was calculated as the total power in the power spectrum from 1 to 8 cycles/image and from 9 to 113 cycles/image, respectively (Le et al., 2016). Each total power of high or low spatial frequency was statistically compared with one-way ANOVA and unpaired *t*-test in the same way as described above. The results of statistical comparisons indicated that none of the statistically significant difference was found in the tests (see Supplementary Tables 1 and 2 for the details).

2.4. Behavioral task

The procedures are detailed in our previous studies (Le et al., 2013; Dinh et al., 2018). Briefly, a 19-inch computer display was placed 68 cm away from a monkey chair for a behavioral task. The monkey's eye position of the left eye was monitored by an eye-monitoring system using a CCD camera (Matsuda, 1996). The timing of visual stimulus onset and juice delivery was controlled by a visual stimulus control unit (ViSaGe MKII Visual Stimulus Generator, Cambridge Research Systems).

The monkeys were required to perform a delayed nonmatching-to-sample task

(DNMS) to ensure that the monkeys looked at visual stimuli not associated with reward in the same condition (i.e., in the central visual field without saccadic eye movements). In the DNMS, the monkeys compared and discriminated the visual stimuli presented in sequence (Fig. 1B). In the task, a fixation cross appeared in the center of the monitor after a buzzer tone. After fixating to the cross for 1.5 s, a sample stimulus appeared for 500 ms (sample phase). The same stimulus appeared again for 500 ms between 1 and 4 times after each 1.5-s interval of fixation. Finally, a new target stimulus appeared (target phase), and the monkeys pressed a button within 2 s to obtain a juice reward (0.8 mL). When the monkeys did not correctly press the button within the 2 s in the target phase, the trials terminated and a buzzer tone (620-Hz) was presented. The trials were repeated with intertrial intervals of 15–25 s.

Thus, the monkeys compared a pair of stimuli (i.e., sample vs. target stimuli) in each trial. The stimuli were randomly selected within the same stimulus category.

2.5. Training and surgery

The monkeys were trained to perform the DNMS task for 3 h/ day, 5 days/week so that performance of the monkeys was asymptotic. All visual stimuli were presented a similar number of times to the monkeys only during training and recording sessions. After 3 months of training, the monkeys reached a 90% correct-response rate. Then, a head-restraining device (U-frame made of reinforced plastic) to receive fake ear-bars was implanted on the skull under anesthesia induced by medetomidine (0.5 mg/kg, i.m.) - ketamine (5 mg/kg, i.m.) combination (Nishijo et al. 1988a, b; Tazumi et al. 2010; Dinh et al., 2018). The U-frame was anchored with dental acrylic to titanium screws twisted into the skull. A small reference pin was also implanted, and its position was calibrated according to the coordinates of the stereotaxic atlas of the *M. fuscata* brain (Kusama and Mabuchi, 1970). Antibiotics (Ceftriaxone sodium, 20 mg/kg) was administered for one week after the surgery. After a recovery period of two weeks after the surgery, the monkeys were retrained with their heads painlessly fixed to the stereotaxic apparatus to reach performance criterion (>90%).

2.6. Neurophysiological recording procedures

The procedures were essentially the same as in earlier studies (Le et al., 2013; Dinh et al., 2018). Briefly, a glass-insulated tungsten microelectrode (0.5-1.5 M Ω at 1 kHz), the tip location of which was calibrated from the reference pin, was stereotaxically inserted into the amygdala with a micromanipulator (Narishige, Tokyo, Japan) according to the stereotaxic atlas of the *M. fuscata* brain (Kusama and Mabuchi, 1970). Neuronal activity with an S/N ratio greater than 3:1 was recorded. All data including neuronal activity, trigger signals of task events, and the coordinates of eye positions were stored in a computer via a data processor (MAP; Plexon, Dallas) system. After recording of responses to the 32 original visual stimuli, the neurons were further tested with the transformed images if the single-unit activity was still observed.

Trials, when eye locations deviated larger than 1.0 degrees from the fixation cross during the fixation and sample periods, were excluded from the analysis. The stored neuronal activities were sorted into single units using the Offline Sorter program for cluster analysis (Plexon). After isolation of clusters, an autocorrelogram for each isolated unit was constructed to observe refractory periods, and units with refractory periods greater than 1.2 ms were selected. Finally, superimposed waveforms of the isolated units throughout the recording sessions were assessed to check variability during recording. These isolated units were further analyzed by the NeuroExplorer program (Nex Technologies, MA, USA) (see below).

2.7. Statistical procedures for analyzing amygdalar neuron responses

The data were analyzed as in two previous studies (Le et al., 2013; Dinh et al., 2018). The data only during the sample phase, but not the target phase, were analyzed. Significant (excitatory or inhibitory) responses to each stimulus were defined by comparison of neuronal activity during the following two periods [Wilcoxon signed-rank (WSR) test, $p < 0.05$]: 100 ms before (pre) and 500 ms after (post) the stimulus onset. Visually responsive neurons were defined as those that responded to one or more visual stimuli (WSR test, $p < 0.05$).

In each responsive neuron, the mean response magnitudes to each stimulus category were computed. Each responsive neuron was categorized by the best category that elicited the largest responses. For example, “snake-best” neurons were those that showed larger mean response magnitude to all snake images than those to the remaining stimulus categories. In each responsive neuron, the mean response magnitudes to each stimulus were computed. Then, the grand mean response magnitudes of the four stimuli in each category were compared among the eight stimulus categories by one-way ANOVA ($p < 0.05$) with Tukey post hoc tests ($p < 0.05$). Furthermore, mean response magnitudes of all responsive neurons ($n = 95$) were compared among the eight stimulus categories by repeated measures one-way ANOVA ($p < 0.05$) with Bonferroni post-hoc tests ($p < 0.05$). In the above analyses of response magnitudes and best-stimulus category, absolute values were used for neurons with inhibitory responses ($n = 4$).

Response latencies to the eight stimulus categories were also analyzed. First, for each neuron, each peri-event histogram for each stimulus category was constructed using the entire set of data for all trials for each stimulus category. Then, neuronal response latency was estimated as the duration between the stimulus onset and the moment at which neuronal activity exceeded the mean \pm 2SD of neuronal activity during the 100-ms pre-period before the stimulus onset. The mean latencies to the eight stimulus categories of all neurons, in which latencies to all eight categories could be estimated using the above criterion ($n = 75$), were compared by repeated measures one-way ANOVA ($p < 0.05$) with Bonferroni post-hoc tests ($p < 0.05$).

To analyze the effects of facial expression (neutral vs. emotional) and species (monkeys vs. humans) on amygdalar neuronal activity, response magnitudes and latencies to the individual monkey and human faces were similarly estimated. Then, mean response magnitudes and latencies ($n = 95$ for response magnitudes; $n = 75$ for response latencies) to the monkey and human faces were analyzed by repeated measures two-way ANOVA ($p < 0.05$) with expression and species as factors.

Finally, response magnitudes to individual stimuli of the snakes, monkey faces, monkey hands, and simple figures were estimated to analyze relationships between the amygdala (present data) and pulvinar (Le et al., 2013) by a linear regression analysis. All data were expressed as mean \pm SEM. In ANOVA statistical analyses, sphericity was assessed, and the degrees of freedom were corrected by Greenhouse-Geisser method where appropriate. All statistical analyses were conducted using SPSS (v. 28, IBM) and GraphPad Prism v. 9.2.0 (GraphPad Software, San Diego, CA).

2.8. Multidimensional Scaling analysis (MDS)

The data were analyzed as in two earlier studies (Le et al., 2013; Dinh et al., 2018). To investigate stimulus representation in the amygdala, data matrices of response magnitudes during specific periods after stimulus onset were analyzed by multidimensional scaling (MDS). MDS creates a geometric representation of stimuli based on the relationship (dissimilarity in the present study) between stimuli to quantitatively analyze similarity of groups of given items (see Hout et al., 2013 for more details). Items are placed in MDS space so that items with similar response patterns are located proximally. Each dimension of the MDS space represents a different underlying factor to estimate similarity.

The data in four 50-ms epochs comprising the initial 200-ms post period were analyzed using MDS. In each 50-ms epoch, the data matrix of response magnitudes in a 95×32 array derived from the 95 visually responsive neurons was generated. The response magnitude in each epoch was computed as the mean firing rate in each epoch subtracted by the mean firing rate during the 100-ms pre-period before the stimulus onset. In MDS, the MDS program (SPSS, v. 28) initially computed dissimilarity (Euclidean distances) between all pairs of two visual stimuli based on the response magnitudes of the 95 amygdalar neurons. Then, the program placed the 32 visual stimuli in the stimulus space so that the distribution of the stimuli in the MDS space represented the original relationships between the stimuli (i.e., Euclidean distances in the present study) (Shepard et al., 1962). Finally, the separation of clusters of the visual stimuli in the stimulus space was assessed by discriminant analyses.

2.9. Localization of the recording sites

The animals were treated as in a previous study (Dinh et al., 2018). Before recording, MRI scans of the monkey head with a tungsten marker (diameter, 800 μm) inserted near the recording region were taken under anesthesia. After the last recording, tungsten markers (diameter, 500 μm) were implanted near the recording sites under anesthesia. Then, the monkeys were anesthetized (sodium pentobarbital, 100 mg/kg, i.m.) and perfused initially with 0.9% saline, and then with 10% buffered formalin. After perfusion, the brains including the amygdala were cut into 100- μm coronal sections, which were stained with Cresyl violet. The locations of the marker tips were determined microscopically, and the location of each recording site was determined by comparing the stereotaxic coordinates of recording sites and those of marker positions.

In the present study, anatomical classification and nomenclature of the amygdalar subnuclei followed the stereotaxic atlas of the *M. fuscata* brain (Kusama and Mabuchi, 1970). According to the previous studies (Nishijo et al., 1988a, b), the amygdala was divided into four areas; CM, corticomедial group of the amygdala (CM), lateral nucleus (AL), basolateral nucleus (Abl), and basomedial nucleus (Abm).

3. Results

3.1. Responses to eight categories of the visual stimuli

Of 1253 amygdala neurons recorded from Monkey1 (M1) ($n = 1151$) and Monkey2 (M2) ($n = 102$), 527 were tested with the eight categories of the non-transformed visual stimuli (M1, $n = 486$; M2, $n = 41$). Of these 527, 95 neurons responded to one or more visual stimuli (visually responsive neurons) (M1, $n = 88$; M2, $n = 7$). Figure 2 shows an amygdalar neuron that responded selectively to snakes. The neuron responded strongly to snakes and less to other stimuli (Fig. 2A). The mean response magnitudes (grand mean responses of mean responses to each stimulus in each stimulus category) of the neuron significantly differed among the stimulus categories [one-way ANOVA: $F(7, 24) = 10.7$, $p < 0.0001$] (Fig. 2B), and the mean response magnitudes to snakes were significantly greater than those to the other stimulus categories including raptors and carnivores (Tukey test, $p < 0.05$), monkey faces (Tukey test, $p = 0.0043$), and non-predators, human faces, simple figures, and monkey hands (Tukey test, $p < 0.0001$). To compare the response trends to the visual stimuli between the two monkeys (Monkey1 and Monkey2), mean response magnitudes were compared among the eight categories in each monkey (Supplementary Figure 1). In Monkey1, response magnitudes to snakes, monkey faces, and human faces were significantly stronger to the other stimulus categories (Supplementary Figure 1A). In Monkey2, a comparable response trend was observed: response magnitudes to snakes and monkey faces were significantly stronger to the other stimulus categories except for human faces

(Supplementary Figure 1B). Furthermore, mean responses to each visual stimulus were significantly and positively correlated (Supplementary Figure 2). These results indicate that the response trends were similar between the two monkeys. Therefore, in the following analyses, the combined data of the two monkeys were analyzed.

The amygdalar neurons were categorized by “best-category” that elicited the largest responses (Le et al., 2013). Of the 95 visually responsive neurons, snake-best neurons were most frequent ($n = 28$), followed by monkey face-best neurons ($n = 26$; 27.4%), and human face-best neurons ($n = 21$; 22.1%). Mean response magnitudes of the 95 visually responsive neurons significantly differed among the eight stimulus categories [repeated measures one-way ANOVA: $F(3.831, 360.1) = 36.05, p < 0.0001$] (Fig. 3A). The mean response magnitudes were significantly stronger to snakes than to the remaining stimuli except for the monkey faces (Bonferroni test, $p < 0.01$). Furthermore, the mean response magnitudes were significantly stronger to the monkey and human faces than to the monkey hands, raptors, carnivores, simple figures, and non-predators (Bonferroni test, $p < 0.01$).

Response latencies of the amygdalar neurons ranged from 40 to 415 ms ($n = 75$). Mean response latencies also significantly differed among the eight categories [repeated measures one-way ANOVA: $F(5.999, 443.9) = 10.80, p < 0.0001$] (Fig. 3B). The mean response latency was significantly shorter for the snakes than for the monkey faces (Bonferroni test, $p < 0.05$), and the human faces, carnivores, raptors, simple figures, non-predators, and monkey hands (Bonferroni test, $p < 0.0001$). Furthermore, the mean response latencies were significantly shorter for the monkey faces than for the raptors, simple figures, non-predators, and monkey hands (Bonferroni test, $p < 0.05$).

The 26 snake-best neurons were further tested with not only the original snake images but also the transformed images including the low-pass filtered snake images, high-pass filtered snake images, Fourier-scrambled snake images, Wavelet-scrambled snake images, and adjusted monkey face images, adjusted human face images, adjusted carnivore images, and adjusted raptor images (Fig. 3C). The mean response magnitudes significantly differed among these nine stimulus groups [repeated measures one-way ANOVA: $F(5.835, 145.9) = 12.92, p < 0.0001$], and the mean response magnitude to the original snakes were significantly greater than those to other stimuli except for the low pass-filtered snake images (Bonferroni test, $p < 0.0001$). Furthermore, the mean response magnitude to the low pass-filtered snake images was significantly larger than those to the high pass-filtered snake images, scrambled snake images (Fourier, Wavelet), and adjusted images of the monkey faces, human faces, carnivores, and raptors (Bonferroni test, $p < 0.05$).

3.2. Responses to emotional faces

Previous studies reported that emotional faces, but not neutral faces, were preferentially processed in the subcortical visual pathway (Vuilleumier et al., 2003; Le et al., 2013; Diano et al., 2017). To investigate this possibility in the monkey amygdala, mean response magnitudes to the monkey and human faces were analyzed with repeated measures two-way ANOVAs with species (monkey vs. human) and expression (emotional vs. neutral) as factors (Fig. 4A). The statistical results indicated significant main effects of species [$F(1.0, 94.0) = 5.810, p = 0.0179$] and expression [$F(1.0, 94.0) = 88.86, p < 0.0001$], but no significant interaction between these two factors [$F(1.0, 94.0) = 2.039, p = 0.1567$]. Mean response latencies to the monkey and human faces were similarly analyzed (Fig. 4B). The statistical results of repeated measures two-way ANOVAs indicated significant main effects of expression [$F(1.0, 74.0) = 89.34, p < 0.0001$] and species [$F(1.0, 74.0) = 5.792, p = 0.0186$], but no significant interaction between the two factors [$F(1.0, 74.0) = 0.1274, p = 0.721$].

3.3. Relationships to the macaque pulvinar neurons

Thus, the amygdalar neurons responded stronger and faster to snakes and emotional faces, which was very similar to the previous study on the macaque pulvinar neurons (Le et al., 2013). Previous anatomical studies reported that the amygdala receives visual information of threatening stimuli from the SC-pulvinar in tree shrews and macaques (Day-Brown et al., 2010; Elorette et al., 2018). If the subcortical visual pathway is involved in processing of innate biological significance of visual stimuli, we hypothesized that activity of amygdalar neurons would be correlated to activity of the pulvinar neurons even in different monkeys. To investigate possible relationships between the amygdala and pulvinar, the correlation between neuronal responses in the pulvinar (Le et al., 2013) and those in the amygdala (present study) was analyzed (Fig. 5), since this and previous studies used the same visual stimuli (a total of 16 stimuli: snakes, monkey faces, monkey hands, and simple figures) tested in the same DNMS task. The results indicated that response magnitudes were significantly and positively correlated between the amygdala and pulvinar [simple regression analysis: $r^2 = 0.820$; $F(1, 14) = 63.80$, $p < 0.0001$].

3.4. Stimulus representation in the amygdala

The four data matrices in epochs 1 (0-50 ms), 2 (50-100 ms), 3 (100-150 ms), and 4 (150-200 ms) after stimulus onset, each consisting of response magnitudes to the 32 visual stimuli in the individual 95 visually responsive amygdalar neurons, were separately analyzed using MDS. In the resultant 2D spaces, r^2 values of epochs 1, 2, 3 and 4 were 0.481, 0.784, 0.618, and 0.730, respectively. In epoch 2 (Fig. 6A), two groups were observed: a cluster containing the snakes (sn1-4) and emotional faces (mf 1-2, hf 1-2), and another containing the remaining stimuli (non-snake and non-emotional face images). These two clusters (snakes and emotional faces vs. non-snakes and non-emotional faces) were significantly separated (discriminant analysis: Wilks' lambda = 0.338, $p < 0.0001$). In epoch 3 (Fig. 6B), three groups were observed; a cluster containing the snakes (sn1-4) and emotional faces (mf 1-2, hf 1-2); a cluster containing the neutral faces (mf 3-4, hf 3-4), and a cluster containing all the other stimuli. These three clusters were also significantly separated (discriminant analysis: Wilks' lambda = 0.155, $p < 0.0001$). These MDS findings indicated that the activity patterns of the population amygdalar neurons discriminated snakes, emotional faces, and neutral faces. However, in epoch 1 (Supplementary Figure 3) and epoch 4 (Supplementary Figure 4), no significant stimulus cluster was observed.

3.5. Locations of the amygdalar neurons

Figure 7A shows the recording sites of responsive neurons in the amygdala. The amygdala was divided into four areas: corticomedial (CM) group of the amygdala, and basolateral (BL) group of the amygdala including the lateral (AL), basolateral (ABl), and basomedial (ABm) nuclei. The ratios of the responsive neurons differed significantly between the corticomedial and basolateral groups (Fig. 7B). The ratios of the visually responsive neurons from all the recorded neurons were significantly greater in the corticomedial group than the basolateral group (χ^2 test: $\chi^2 = 11.87$, $p = 0.0006$). Furthermore, ratios of the neurons that responded significantly to snakes from all the recorded neurons were significantly larger in the corticomedial group than the basolateral group (χ^2 test: $\chi^2 = 15.26$, $p < 0.0001$). In addition, ratios of the neurons that responded significantly to monkey faces from all the recorded neurons were significantly larger in the corticomedial group than the basolateral group (χ^2 test: $\chi^2 = 6.895$, $p = 0.0086$).

4. Discussion

In the present study, I found that amygdalar neurons responded selectively to snake and face stimuli: (i) mean response magnitudes for snakes, monkey faces, and human faces were greater than those for other stimuli; (ii) mean response latencies for snakes and monkey faces were shorter than for other stimuli; and (iii) activity patterns of the 95 amygdalar responsive neurons discriminated snakes and emotional faces within a 100-ms latency. These responses to snakes were dependent at least on low-frequency components of the images, while high-pass filtering and scrambling of the snake images decreased neuronal responses. The present results provide neurophysiological evidence that amygdalar neurons receive fast and coarse visual information of fear-relevant stimuli (snakes and emotional faces), which may be effective in eliciting rapid behavioral responses in response to those stimuli in nature (see below).

4.1. Responsiveness of amygdalar neurons

Consistent with studies of the “fear module” where snakes and emotional faces are particularly salient stimuli in humans (Morris et al., 2001; Almeida et al., 2015), the present study indicated greater responsiveness of amygdalar neurons to snakes and emotional faces than to other predators (carnivores and raptors) and neutral stimuli. Fourier- and Wavelet-scrambling of the snake images decreased responses to the snakes. In addition, responses to the adjusted images of the monkey and human faces, carnivores, and raptors, in which low-level properties of the images were equalized with means of the low-level properties of the snake images, were smaller than those to the original snake images. These findings suggest that the high responsiveness to snakes is less dependent on global low-level properties of the original snake stimuli, but more dependent on the form of the snakes.

High spatial, but not low spatial, filtering of snake images decreased response magnitudes to the snakes, suggesting that preferential responsiveness to the snake images was dependent on its low spatial frequency component in the present study. Consistently, high spatial, but not low spatial, filtering of snake images decreased response magnitudes to the snakes in the monkey pulvinar that projects to the amygdala (Le et al., 2013). Furthermore, low spatial frequency images of emotional faces elicited fast synaptic responses in the human amygdala in early latencies (less than 100 ms) before cortical responses (Méndez-Bértolo et al., 2016). Human fMRI studies are also consistent with the present results: low spatial component of emotional faces elicited greater responses in the amygdala than the low spatial component of neutral faces in a patient with V1 lesions and intact humans (Vuilleumier et al., 2003; Burra et al., 2019). These findings suggest that the amygdala receives crude features of dangerous visual signals faster than the visual cortex and consequently is involved in coarse and fast processing of such signals, which might be crucial for self-defense and survival.

4.2. Detection of snakes and emotional faces in the amygdala

Previous behavioral studies consistently reported that amygdalar lesions decreased aversive behavioral responses to snakes in monkeys and human patients (Meunier et al., 1999; Kalin et al., 2001; Izquierdo et al., 2005; Mason et al., 2006; Machado et al., 2009; Feinstein et al., 2011), and disturbed recognition of emotional faces in human patients (Adolphs et al., 1994; Calder et al., 1996). Our MDS results indicated that snakes and emotional faces were separated from the other stimuli in the second epoch (50-100 ms after stimulus onset), suggesting a pivotal role of the amygdala in fast detection of emotional stimuli. This early time window strongly suggests that the amygdala directly receives visual inputs from the subcortical pathway bypassing the cortical routes. In contrast, neutral faces were separated in the third epoch (100-150 ms after stimulus onset). This suggests that cortical inputs such as those from temporal face areas may also contribute to amygdalar neuronal responses to facial stimuli (Pitcher et al., 2017).

A recent monkey behavioral study reported that bilateral selective lesions of the central nucleus of the amygdala also disturbed aversive behavioral responses to snakes (Kalin et al., 2004). Consistent with this finding, most human fMRI studies reported that emotional stimuli activated the amygdala, especially its dorsal or dorsomedial part (Breiter et al., 1996; de Gelder et al., 2005; Schaefer et al., 2014; Hakamata et al., 2016), which roughly corresponds to the corticomedial group of the amygdala that includes the central, medial, and cortical nuclei of the amygdala. The amygdala is functionally divided into two regions with reciprocal connections (Aggleton, 1985; Bielski et al., 2021): the dorsomedial part that corresponds to the corticomedial group of the amygdala (output nuclei for behavioral responses), and the ventrolateral part that corresponds to the basolateral group of the amygdala (affective evaluation of sensory stimuli). Consistent with the above fMRI studies, the present study indicated that the ratios of the responsive neurons were significantly larger in the corticomedial group than the basolateral group of the amygdala. This suggests that the corticomedial group of the amygdala is more responsive to various types of aversive stimuli, and consequently, it may be involved with stereotyped first response, subconscious, subcortically controlled behaviors (e.g., avoidance, freezing, etc.), whereas the basolateral group may be more involved with later, cortically controlled evaluation and decision-making (McGarry and Carter, 2017; Ishikawa et al., 2020).

In the present study, all visual stimuli were similarly presented to the monkeys only during training and recording sessions. Therefore, the differences in responses among the eight categories are likely to be attributed to their innate biological significance, consistent with previous studies reporting that behavioral responses to snakes and faces are innate in monkeys (for snakes: Nelson et al., 2003; Kawai and Koda, 2016; Bertels et al., 2020; for faces: Sackett, 1966; Sugita, 2008). However, previous studies also reported that behavioral responses to these stimuli may be altered by prior experience: macaque monkeys developed a fear of snakes by observational learning (Cook and Mineka, 1990), and stressful early life experience of parents of subjects might alter coping styles of the subjects in response to snakes in marmosets (Clara et al., 2008). Although they did not go through such stressful procedures before or during the present study, they interacted directly with their conspecifics in the breeding facility and encountered humans during training and recording. Furthermore, I could not exclude the possibility that the monkeys might have encountered real snakes at the breeding facility. However, the amygdalar responses to the stimuli in the present study were significantly correlated to the pulvinar responses in the previous study in which the monkeys were unlikely to have encountered real snakes (Le et al., 2013). These findings suggest that amygdalar responses to snakes were attributed to their innate biological significance.

However, there are several limitations in the present study. First, this study was designed to analyze effects of innate biological significance (biological saliency) of the static visual stimuli, but not stimulus saliency of low-grade visual properties (physical saliency, i.e., luminance, motion, and color of the stimulus). It has been reported that these physical properties of visual stimuli are also critical as stimulus saliency in patients with V1 lesions (Itti and Koch, 2001; Yoshida et al., 2012), suggesting that the subcortical visual pathway is sensitive to such stimuli. Therefore, stimuli such as onset/offset of light with different intensities could affect amygdalar neuronal activity, and the present results do not deny such possibility. I speculate that the subcortical visual pathway may be sensitive to both biological and physical saliency of visual stimuli. Second, I tested amygdalar neurons only with a limited number of static visual images. The results of scrambling the snake images only indicate that stimulus coherency is important for neuronal sensitivity to the snake images. Thus, specific features of the snake images that activate amygdalar neurons were not analyzed in detail. In future studies, it is interesting to compare neuronal activity between

sinusoidal line moving and a snake slither pattern, and to test snake-responsive neurons with a branch of similar thickness and color pattern and gaussian light onset /offset, etc.

4.3. Role of the subcortical pathway in primates

The above discussion suggests that amygdalar neuronal responsiveness is similar to that in the pulvinar and medial prefrontal cortex in monkeys (Major et al., 2010; Le et al., 2013; Dinh et al., 2018). This specific pattern of responsiveness to snakes and emotional faces might be attributed to natural selection favoring rapid detection of snakes as well as emotional faces in primates via the “fear module”, which includes the superior colliculus, pulvinar, and amygdala (Isbell, 2006, 2009; Öhman et al., 2012).

Snakes were the first modern predators of primates (and other mammals), having evolved before carnivorans and raptors, and, because many snakes are sit-and-wait predators, they can often be avoided by initially freezing or jumping away, suggesting that snakes may have been a main driver behind subcortical visual evolution (Isbell, 2006, 2009). In the present study, response magnitudes were positively correlated between the pulvinar and amygdala, consistent with a human fMRI study which reported that activity in the amygdala and pulvinar was positively correlated (Morris et al., 2001; Troiani et al., 2014), suggesting that this subcortical pathway is functional in intact monkeys. A monkey anatomical study also indicated that amygdalar neurons receive visual information from the superior colliculus through the medial part of the pulvinar, where snake-sensitive neurons were most common (Le et al., 2013), and which may contribute to rapid processing of emotional stimuli (Elorette et al., 2018). Furthermore, bilateral lesions of the superior colliculus abolished aversive behavioral responses to snakes in monkeys (Maier et al., 2011).

Almost all anthropoid primates live in groups and are highly social. Flexible facial expressions can help individuals determine the intentions of others (Burrows, 2008; Dobson, 2009; Dobson and Sherwood, 2011), which can range from highly affiliative to highly agonistic, the latter sometimes resulting in death (Martínez-Íñigo et al., 2021). Thus, responding rapidly to avoid a threatening conspecific may reduce injury and increase the chances of survival. As with snakes, emotional faces may have provided a reliable cue for avoiding danger and selective pressure for visual evolution (Soares et al., 2017).

Pulvinar lesions also disturbed unconscious processing of emotional stimuli in human patients (Bertini et al., 2018). In addition, fMRI studies reported that fiber density connecting the superior colliculus, pulvinar, and amygdala, detected using tractography, was positively correlated with functional connectivity between the pulvinar and amygdala, and positively correlated with recognition of emotional faces and attentional bias to threatening stimuli (Koller et al., 2019; McFadyen et al., 2019). These findings suggest that the amygdala’s role in behavioral responses to snakes and emotional faces in primates is dependent on visual inputs to the amygdala through the visual pathway of the subcortical “fear module”. The present results provide complementary neurophysiological evidence of the same visual dependency.

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Figure legends

Fig. 1. Visual stimuli (A) and delayed nonmatching-to-sample (DNMS) task (B).

(Aa) Non-transformed original stimuli. Thirty-two photos of eight categories of the visual stimuli including snakes, monkey faces (emotional and neutral faces), human faces (emotional and neutral faces), monkey hands, non-predators, raptors, carnivores, and simple figures. (Ab) Transformed stimuli. LPF, low-pass filtered images; HPF, high-pass filtered images; Fourier, Fourier-scrambled images; Wavelet, wavelet-scrambled images; Adjusted, adjusted visual images. (B) Schematic illustration of the DNMS task sequence. The photos are used on the courtesy of Mr. D. Hillman (sn 1), Mr. I. Hoshino (sn 2-4), Mr. Rich Lindie (ra 3), Mr. Prasanna De Alwis (ra 4), EarthWatch Institute (ca 2), Microsoft Windows98 (ca 4); "Brahminy Kite juvenile" (ra 2) by Challiyan is licensed under CC BY-SA 3.0; "cow" (np1) and "chicken" (np 2) are freely available pictures from Pixabay; "sheep" (np 3) was purchased from Pixabay; "The black rat" (np 4) by CSIRO is licensed under CC BY-SA 3.0.

Human facial pictures (hf 2, hf 4) were purchased from ATR-Promotions (rights holder of the pictures).

Fig. 2. Neuronal responses of a snake-responsive amygdalar neuron.

(A) Neuronal responses to each indicated stimulus are shown by raster displays and their summed histograms. Horizontal red bars above the raster displays, stimulus presentation periods (500 ms); bin width of histograms, 50 ms; calibration at the right bottom of the figure, number of spikes per trial in each bin. (B) Comparison of response magnitudes of the neuron shown in Fig. 2A to the eight categories of the visual stimuli. ****, **, *, $p < 0.0001$, 0.01, 0.05, respectively (Tukey test after one-way ANOVA).

Fig. 3. Response characteristics of the amygdala neurons to the eight categories of the visual stimuli (A, B) and the transformed stimuli (C).

(A) Comparison of the mean response magnitudes to the eight stimulus categories. **, $p < 0.01$, respectively (Bonferroni test after repeated measures one-way ANOVA). (B) Comparison of the mean response latencies to the eight stimulus categories. ****, *, $p < 0.0001$, 0.05, respectively (Bonferroni test after repeated measures one-way ANOVA). (C) Comparison of averaged response magnitudes of 26 amygdalar neurons to the original and transformed images. Original, original snake images; LPF, low pass-filtered snake images; HPF, high pass-filtered snake images; Fourier, Fourier-scrambled snake images; Wavelet, Wavelet-scrambled snake images; Adjusted, adjusted images. ****, *, $p < 0.0001$, 0.05, respectively (Bonferroni test).

Fig. 4. Effects of expression and species of monkey and human faces on amygdalar neuronal responses.

(A) Comparison of the mean response magnitudes. ****, $p < 0.0001$ (significant main effect of expression: emotional vs. neutral); #, $p < 0.05$ (significant main effect of species: monkey vs. human). (B) comparison of the mean response latencies. ****, $p < 0.0001$ (significant main effect of expression: emotional vs. neutral); #, $p < 0.05$ (significant main effect of species: monkey vs. human).

Fig. 5. Linear correlation of response magnitudes between the pulvinar and amygdala.

Response magnitudes to the same 16 stimuli in the amygdala and those in the pulvinar (Le et al. 2013) are plotted. There was a significant positive correlation between the amygdala and pulvinar in response magnitudes ($p < 0.0001$).

Fig. 6. Representation of the 32 visual stimuli in a 2D space resulting from MDS using responses of the 95 amygdalar neurons to these stimuli.

(A) Representation in epoch 2. The snakes (sn1-4) and emotional faces (mf 1-2, and hf 1-2) were separated from the remaining stimuli. (B) Representation in epoch 3. Three groups were separated: group 1 (snakes and emotional faces: sn1-4, mf 1-2, and hf 1-2), group 2 (neutral faces: mf 3-4, and hf 3-4), and group 3 containing the remaining stimuli.

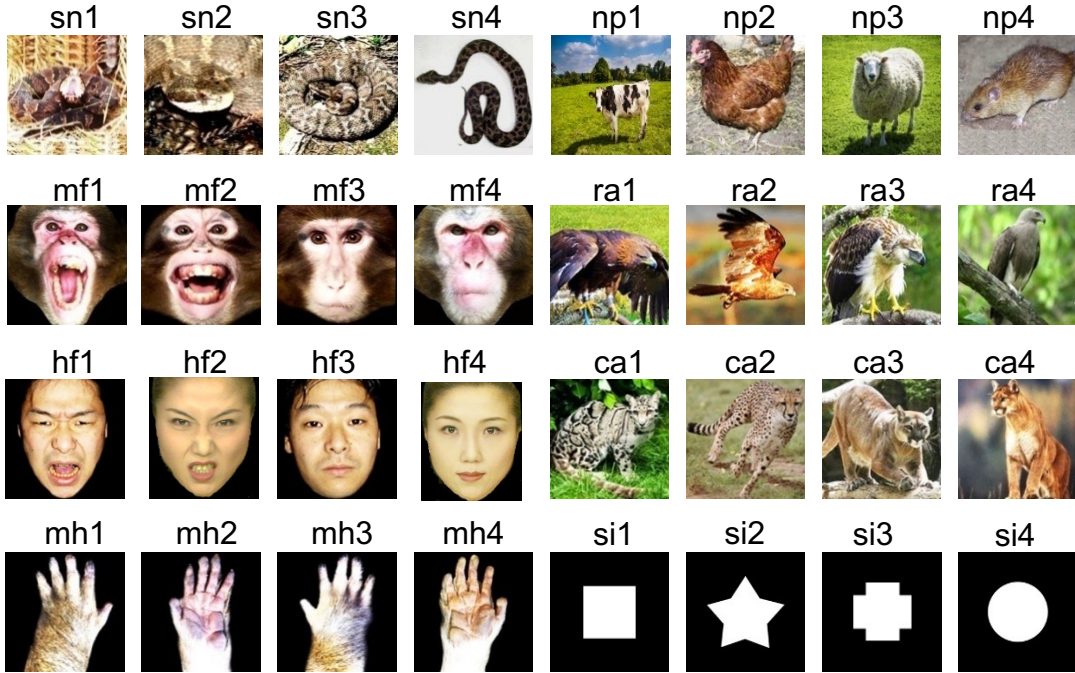
Fig. 7. Recording sites in the amygdala.

(A) Recording sites are plotted on coronal sections at the different anteroposterior axis (A-P levels). Numbers below coronal sections, distance (mm) anteriorly from the interaural line; open circles, responsive neurons ($n = 95$); dots, non-responsive neurons. The amygdala was divided into four areas: CM, corticomедial group; AL, lateral nucleus; Abl, basolateral nucleus; Abm, basomedial nucleus. (B) Ratios of responsive neurons from all the recorded neurons in the corticomедial and basolateral groups of the amygdala. The ratios of the

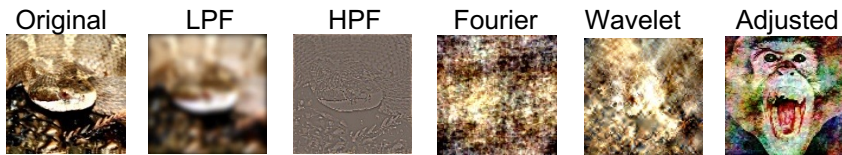
visually responsive (a), snake-responsive (b), and monkey face-responsive (c) neurons were significantly greater in the corticomедial group of the amygdala than in the basolateral group of the amygdala. ****, ***, **, $p < 0.0001, 0.001, 0.01$ (χ^2 test).

A. Visual stimuli

a. Original



b. Transformed



B. Behavioral task

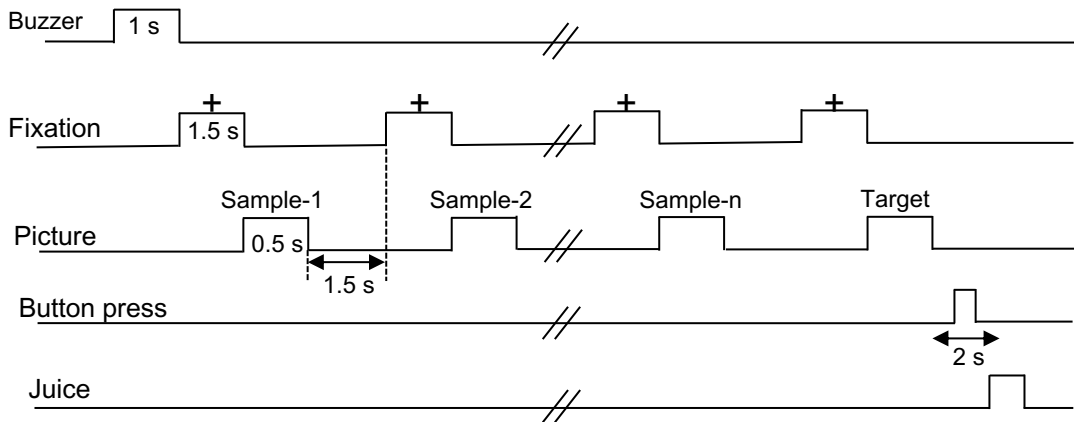


Fig. 1

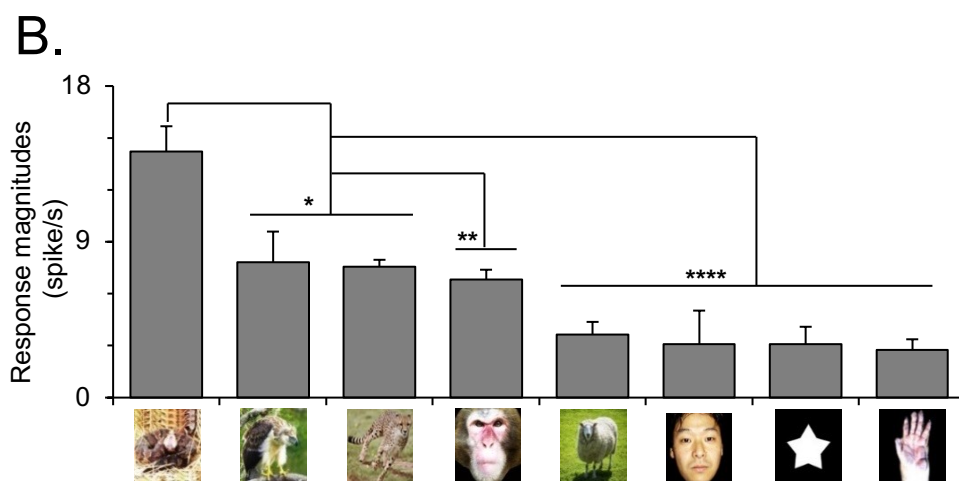
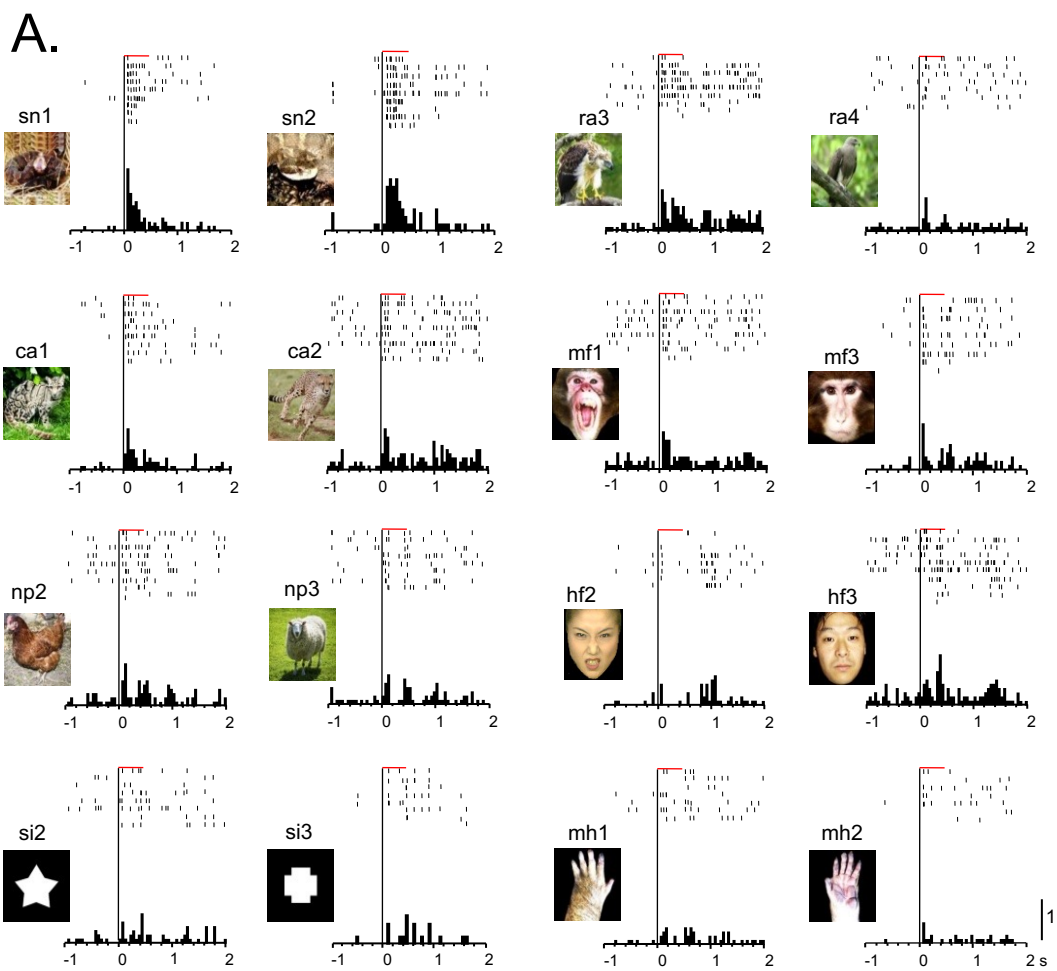
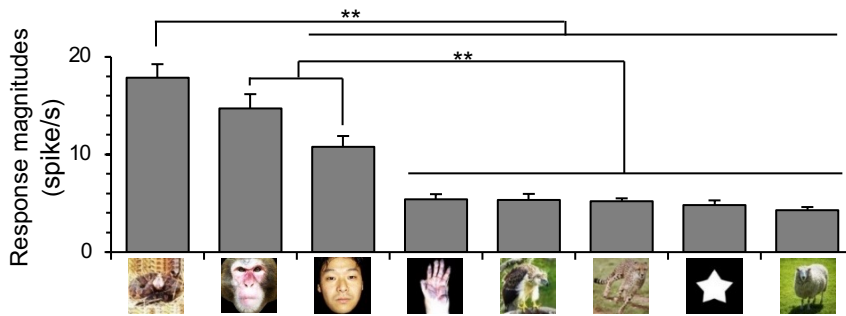
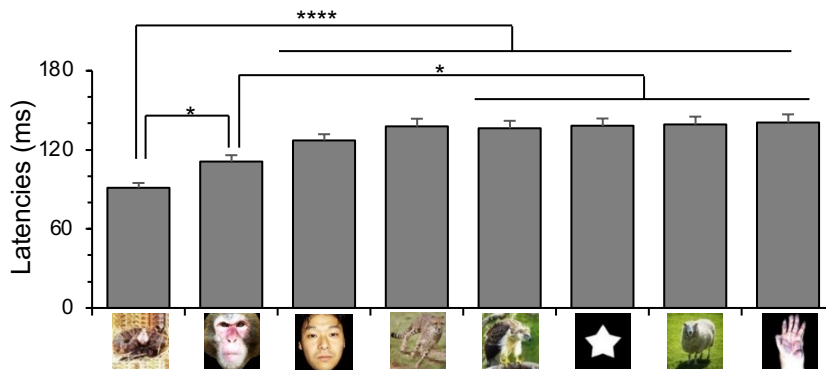


Fig. 2

A. Response magnitude



B. Response latency



C. Transformed stimuli

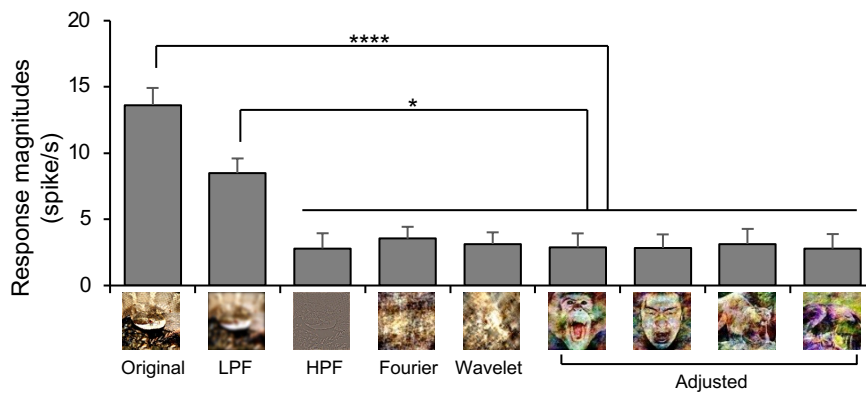
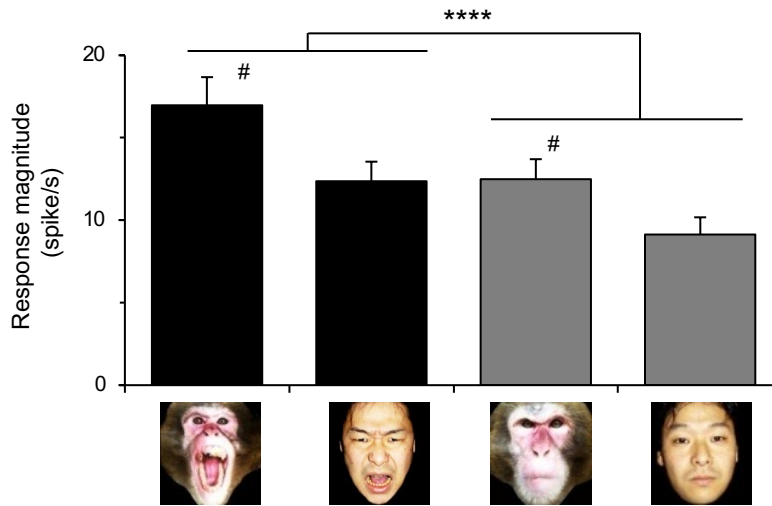


Fig. 3

A. Response magnitudes



B. Response latencies

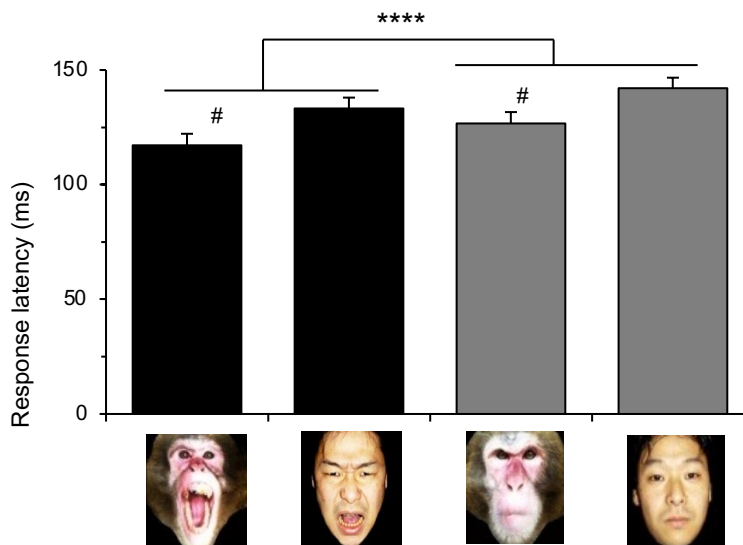


Fig.4

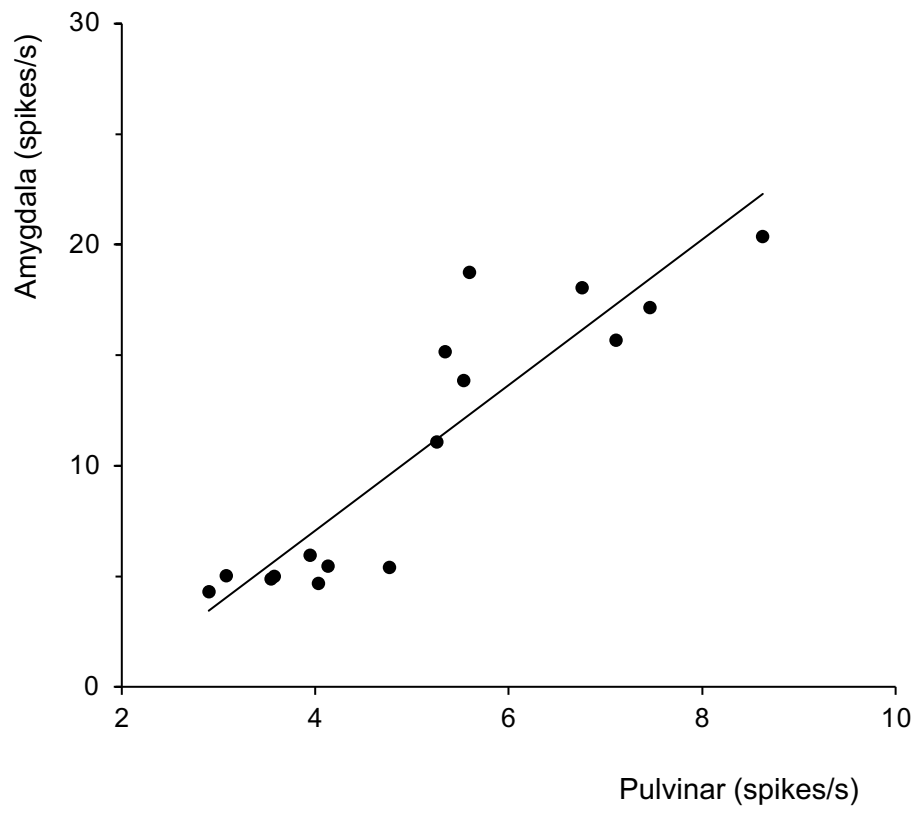
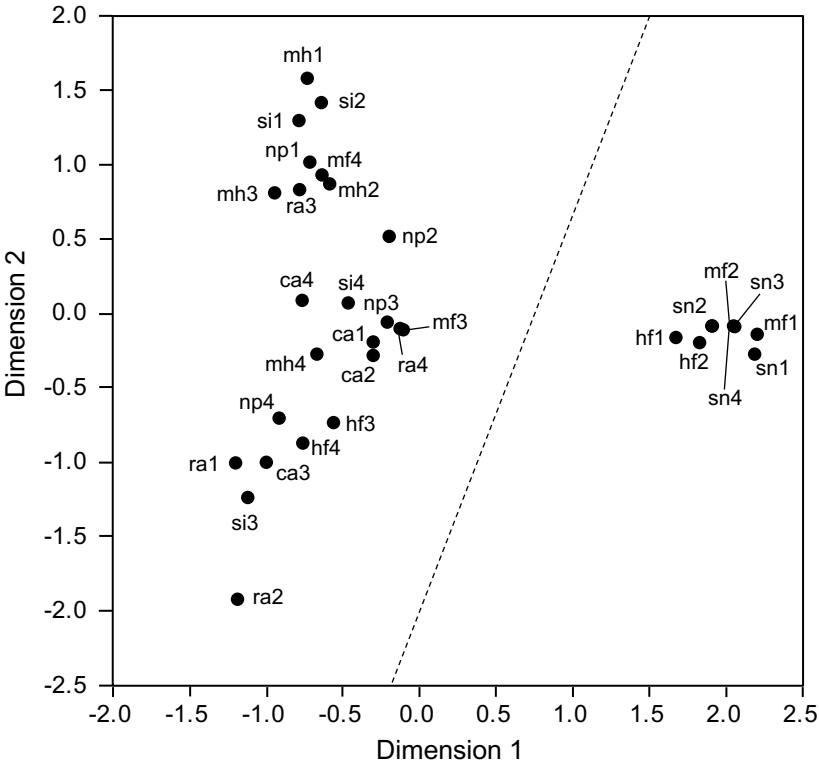


Fig. 5

A. Epoch 2 (50-100ms)



B. Epoch 3 (100-150ms)

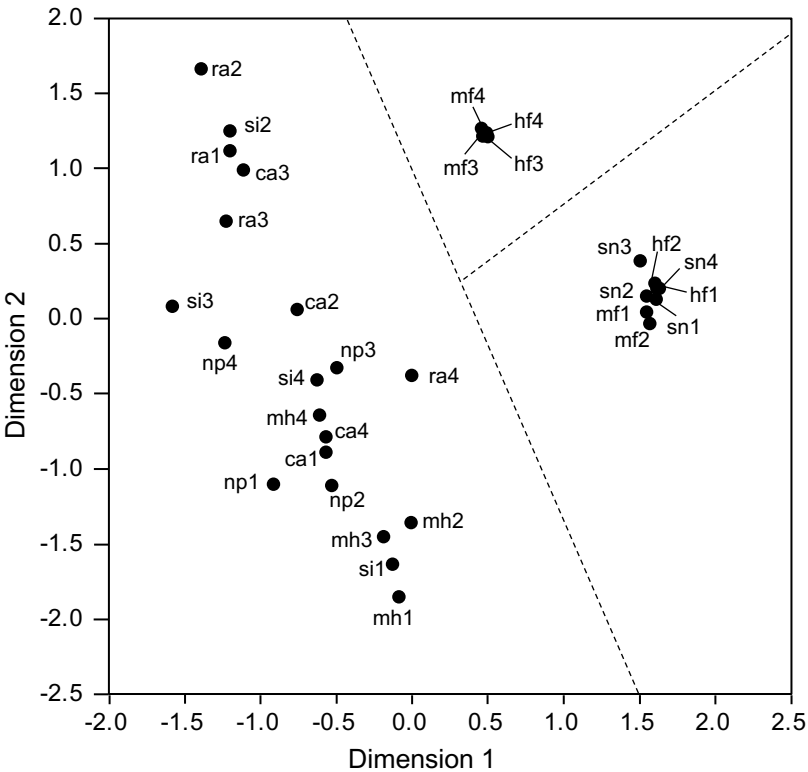
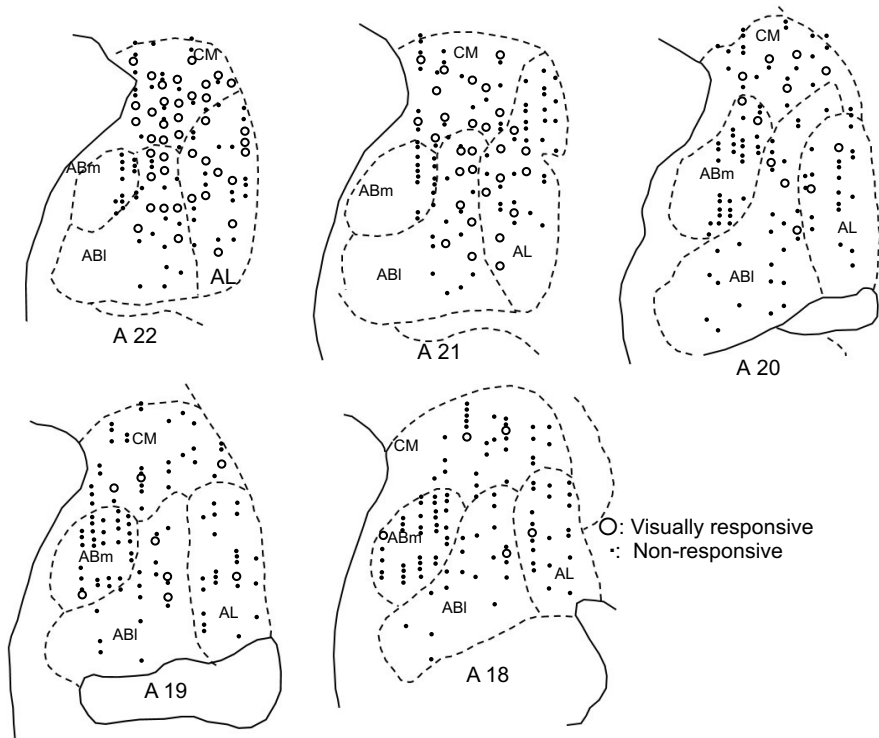


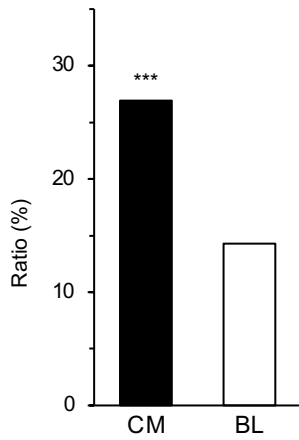
Fig. 6

A.

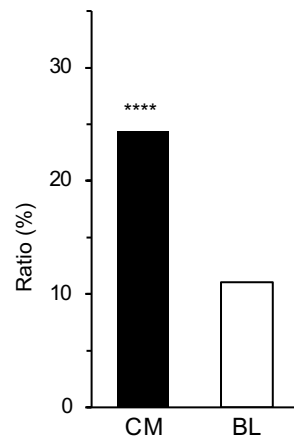


B. Ratios of specific amygdalar neurons

a. Visually responsive



b. Snake-responsive



c. Monkey face-responsive

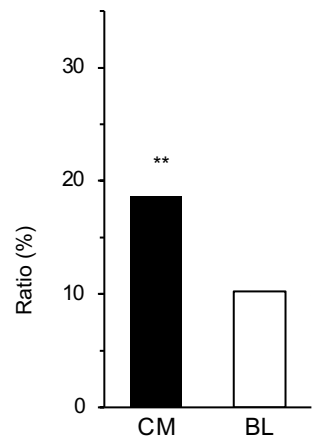


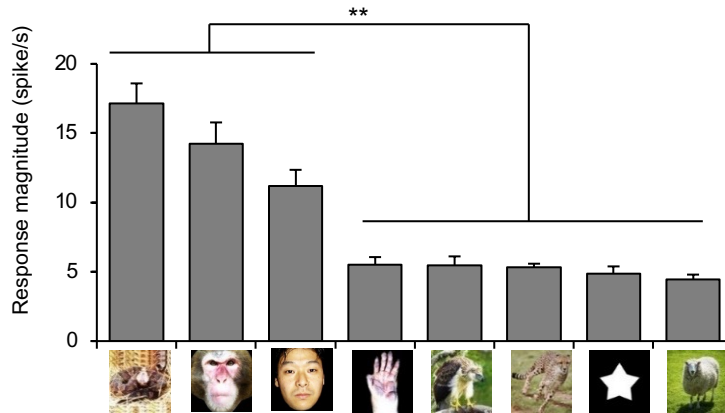
Fig. 7

Fast detection of snakes and emotional faces in the monkey amygdala

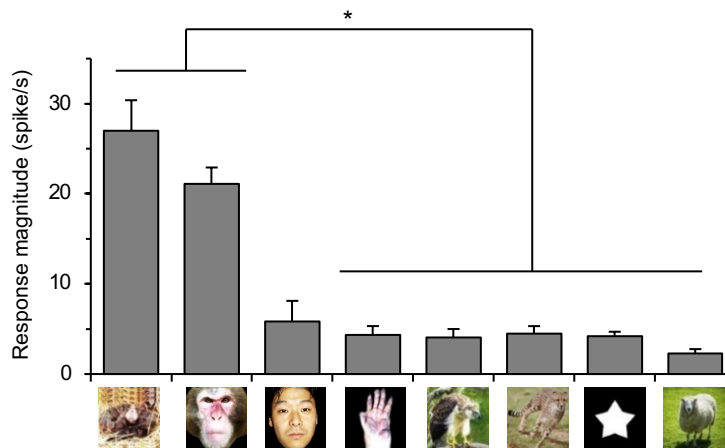
Yang Meng

Supplementary information

A. Response magnitude (M1)

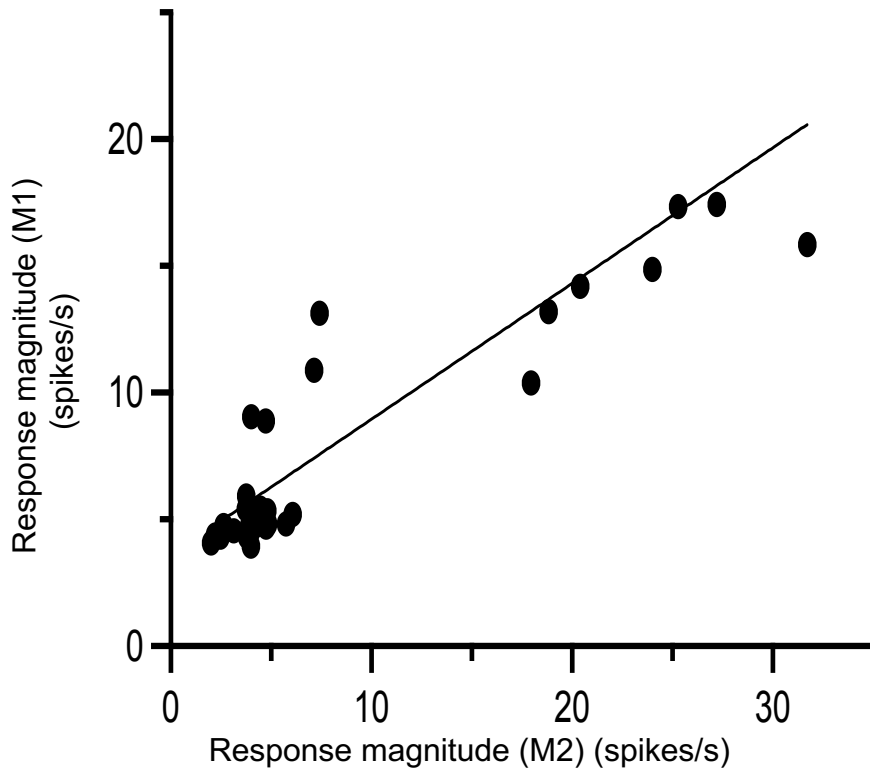


B. Response magnitude (M2)



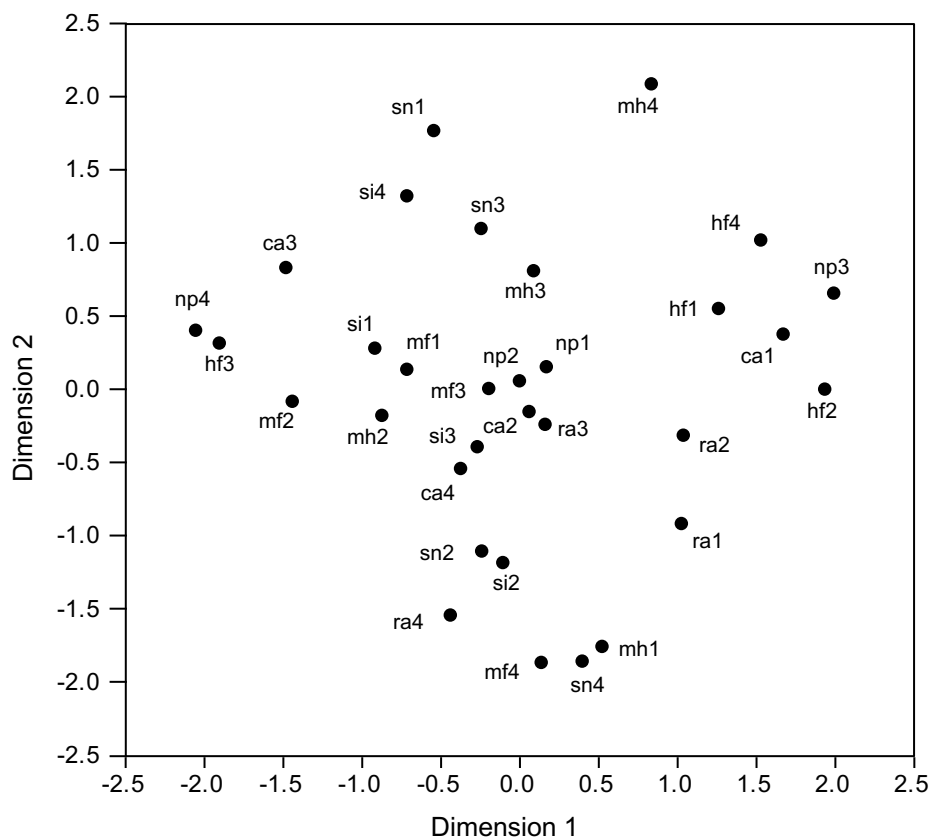
Supplementary Figure 1. Comparison of response magnitudes among the eight categories of the visual stimuli in each monkey.

(A) Comparison of response magnitudes in Monkey1 (M1). There was a significant main effect of stimulus category [repeated measures one-way ANOVA: $F(3.891, 338.5) = 28.87, p < 0.0001$]. **, $p < 0.01$ (Bonferroni test). (B) Comparison of response magnitudes in Monkey2 (M2). There was a significant main effect of stimulus category [repeated measures one-way ANOVA: $F(1.966, 11.79) = 29.30, p < 0.0001$]. *, $p < 0.05$ (Bonferroni test).

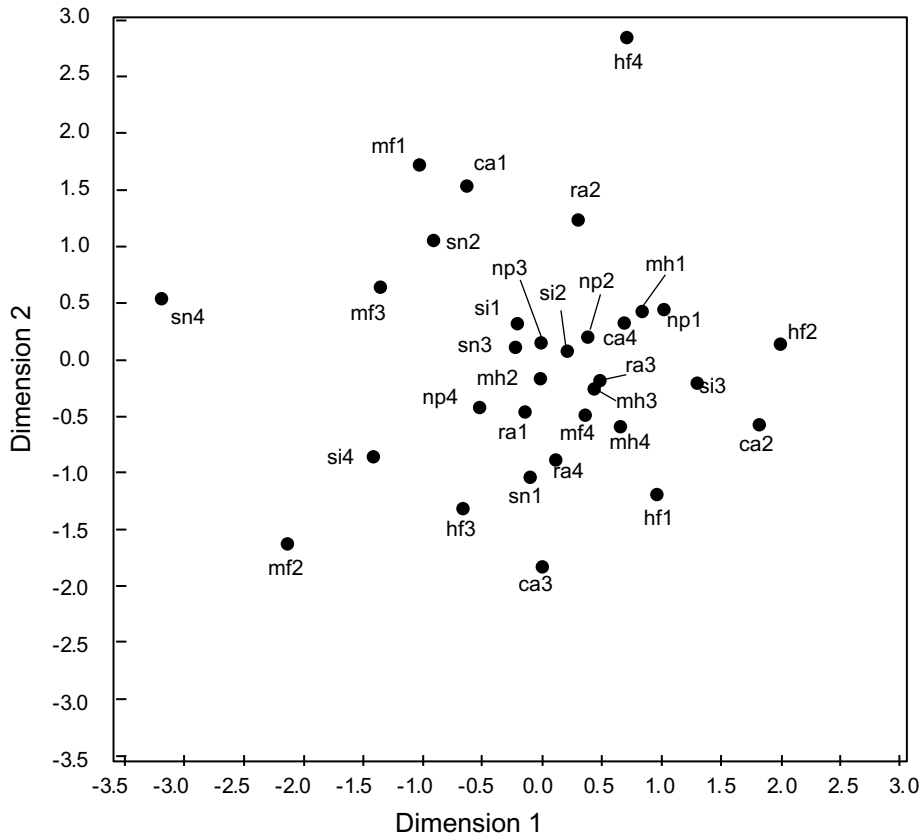


Supplementary Figure 2. Linear correlation of response magnitudes between the Monkey1 (M1) and Monkey2 (M2).

Response magnitudes to the same 32 stimuli in the amygdala of the two monkeys are plotted. There was a significant positive correlation between the two monkeys [$F(1, 30) = 7.102, p < 0.0001$].



Supplementary Figure 3. Distributions of the 32 visual stimuli in a 2D space resulting from MDS using responses of the 95 amygdalar neurons to these stimuli in epoch 1. No stimulus separation was observed. For abbreviations of the stimuli, see Fig. 1Aa.



Supplementary Figure 4. Distributions of the 32 visual stimuli in a 2D space resulting from MDS using responses of the 95 amygdalar neurons to these stimuli in epoch 4. No stimulus separation was observed. For abbreviations of the stimuli, see Fig. 1Aa.

Fast detection of snakes and emotional faces in the monkey amygdala

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Supplementary information (tables)

Supplementary Table 1. Results of one-way ANOVAs among the five categories (original snakes, adjusted raptors, adjusted human faces, adjusted monkey faces, and adjusted carnivores).

			ANOVA results	
			F(4,15)-values	p-values
Color histograms	Red	bin: 0.00-0.25	0.236249856	0.913518009
		bin: 0.25-0.50	0.728040545	0.586543705
		bin: 0.50-0.75	0.030198612	0.998035064
		bin: 0.75-1.00	0.011916594	0.999684519
	Green	bin: 0.00-0.25	0.554500726	0.698921369
		bin: 0.25-0.50	0.098319789	0.981379682
		bin: 0.50-0.75	0.002875815	0.999981345
		bin: 0.75-1.00	0.00412696	0.999961663
	Blue	bin: 0.00-0.25	1.426459565	0.273317008
		bin: 0.25-0.50	0.49406085	0.740340734
		bin: 0.50-0.75	0.041578221	0.996345049
		bin: 0.75-1.00	0.124869561	0.971225194
Michelson contrast		1	0.438000811	
Power spectrum	Low freq. (1-8)	Red	1.801000244	0.181203957
		Green	1.953911767	0.153640659
		Blue	1.145773908	0.373022713
	High freq. (9-113)	Red	0.433822576	0.78214687
		Green	0.447238394	0.772819971
		Blue	0.397795637	0.807148562

Supplementary Table 2. Results (p-values) of t-tests (original snakes vs. adjusted images).

			Categories of the adjusted images			
			Adjusted raptors	Adjusted human faces	Adjusted monkey faces	Adjusted carnivores
Color histogram	Red	bin: 0.00-0.25	0.662	0.603	0.663	0.663
		bin: 0.25-0.50	0.420	0.415	0.452	0.401
		bin: 0.50-0.75	0.876	0.940	0.916	0.847
		bin: 0.75-1.00	0.994	0.936	0.969	0.967
	Green	bin: 0.00-0.25	0.502	0.450	0.491	0.496
		bin: 0.25-0.50	0.793	0.763	0.742	0.774
		bin: 0.50-0.75	0.996	0.995	0.956	0.976
		bin: 0.75-1.00	0.955	0.943	0.977	0.975
	Blue	bin: 0.00-0.25	0.254	0.239	0.312	0.306
		bin: 0.25-0.50	0.451	0.526	0.582	0.524
		bin: 0.50-0.75	0.830	0.967	0.935	0.872
		bin: 0.75-1.00	0.932	0.771	0.949	0.948
Michelson contrast		0.356	0.356	0.356	0.356	
Power spectrum	Low freq. (1-8)	Red	0.230	0.226	0.224	0.232
		Green	0.212	0.207	0.210	0.219
		Blue	0.337	0.316	0.318	0.333
	High freq. (9-113)	Red	0.518	0.600	0.563	0.497
		Green	0.517	0.580	0.544	0.498
		Blue	0.550	0.679	0.571	0.508