

## Facile Preparation of Dehydrodigallic Acid and Its Derivative for the Synthesis of Ellagitannins

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Received June 10, 2013; accepted September 17, 2013

**A facile method for the synthesis of dehydrodigallic acid, which is a fundamental structure of ellagitannins, was developed. A classical Ullmann condition was effective for the formation of the highly hindered biaryl ether structure, and we clarified that the suitable protection of the phenolic hydroxy groups was crucial in this reaction. In this way, the synthesis of dehydrodigallic acid and its derivative was successfully performed. The described method would provide a synthetic utility toward ellagitannins.**

**Key words** ellagitannin; Ullmann reaction; biaryl ether

Ellagitannins are representative polyphenolic secondary metabolites of higher plants and possess a wide range of biological activities (*e.g.*, antioxidant, antiviral, and antitumor activities).<sup>1)</sup> Although this class of compounds is composed only of gallic acid (**1**), oxidized gallic acid, and sugar, a few hundred congeners have been discovered in nature as the mono- or oligomeric form (Fig. 1). Based on their multifunctional property and structural diversity, these compounds have attracted a great deal of attention from many synthetic chemists. One of the most outstanding features of the ellagitannins is furnishing an axially chiral biaryl unit which is formed by the C–C oxidative phenolic coupling of gallic acid (**1**), hence they produce ellagic acid (**2**) upon hydrolysis. From this point of view, synthetic studies of the ellagitannins have focused on the construction of the biaryl part with an axial chirality<sup>2–5)</sup>; however, gallic acid (**1**) also provides dehydrodigallic acid (**3**)<sup>6)</sup> as a skeletal motif of the ellagitannins *via* the biogenetic C–O oxidative coupling reaction in plants. In fact, numerous ellagitannins incorporate this compound as the dehydrodigalloyl (DHDG) group in their structure, such as coriariin B.<sup>7)</sup> Therefore, a convenient method for preparing **3** has been required to

synthesize a diverse set of ellagitannins.

The synthesis of **3** was already accomplished by the classical Ullmann reaction of phenol and an aryl halide that was derived from **1**.<sup>8)</sup> In the C–O bond formation step, however, the yield was very low. In 1996, Feldman and co-workers overcame the difficulty of the construction of the highly functionalized biaryl ether structure of **3** in a completely different way.<sup>9)</sup> They investigated the reactivity of galloyl-derived orthoquinones, such as **4**, and applied it to the hetero Diels–Alder reaction (Chart 1). The four-step sequence involving the Diels–Alder dimerization/reductive rearrangement was moderately effective for the synthesis of **5a** that was the benzylated derivative of **3**, which was the key synthetic intermediate of coriariin A.<sup>10–12)</sup> Although their strategy is very effective for the ellagitannin synthesis, it might be difficult that the two ester groups of **5a** would be distinguished.<sup>13)</sup> This concern must be solved in order to synthesize coriariin B and other ellagitannins. In this context, we thus reinvestigated the synthesis of **3** and its derivative using the Ullmann condensation<sup>14)</sup> that seemed to be convergent, flexible, and efficient, as shown in Chart 1.<sup>15–18)</sup>

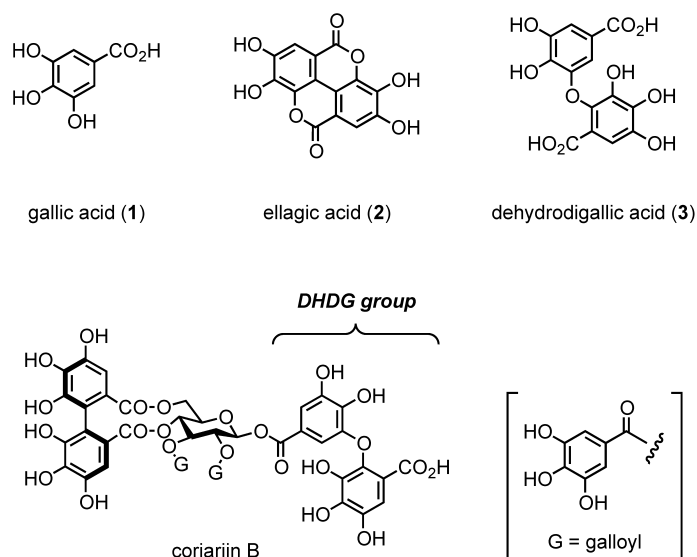


Fig. 1. Structure of Ellagitannin-Related Compounds

The authors declare no conflict of interest.

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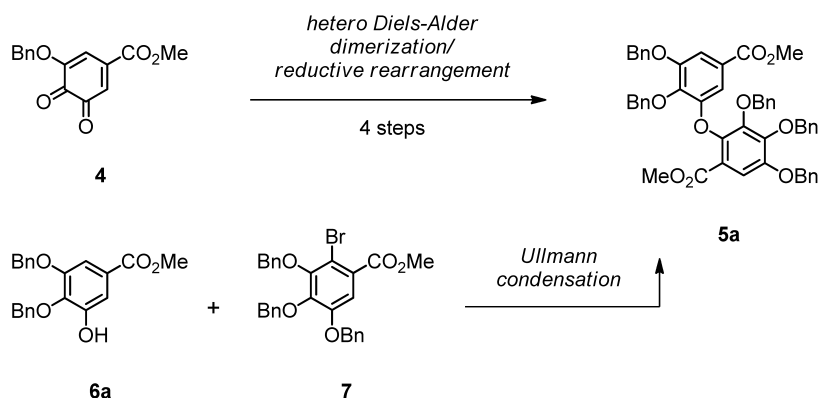
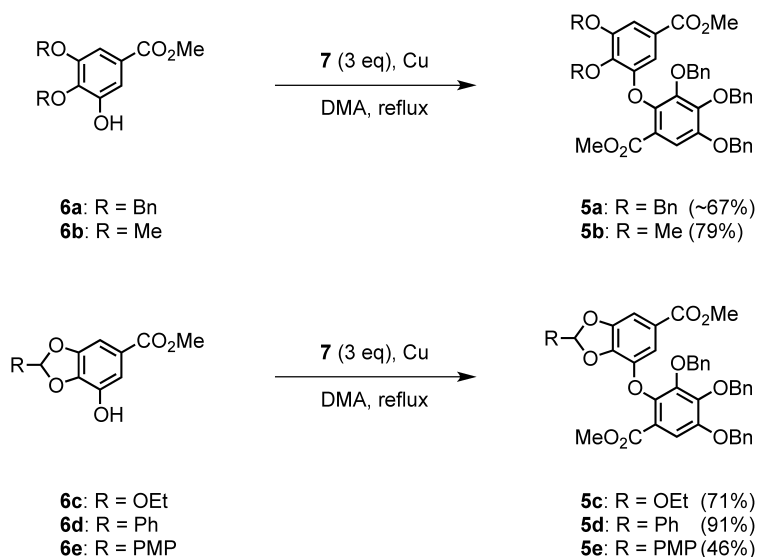
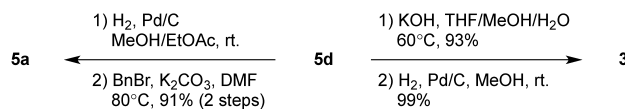


Chart 1. Two Different Approaches for Highly Functionalized Biaryl Ether

Chart 2. Ullmann Reaction of **6a–e** with **7**

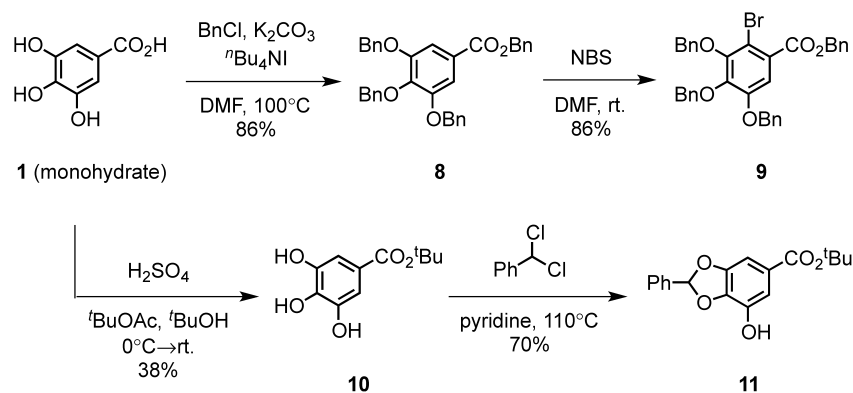
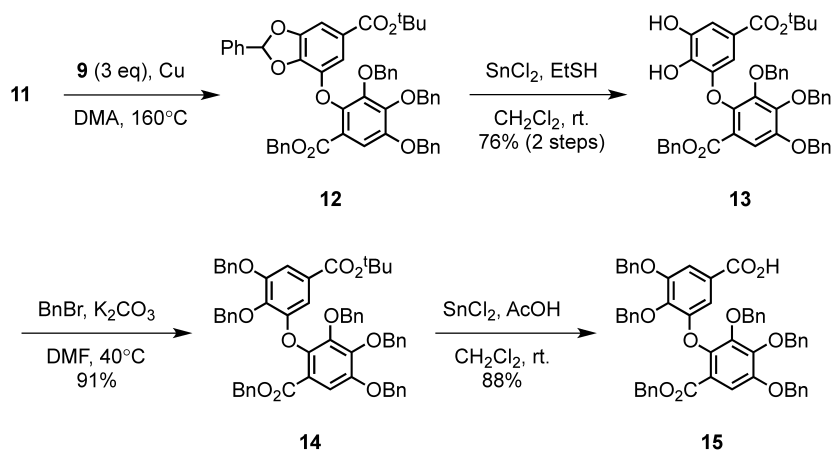
For our initial explorations, the reaction of phenol **6a**<sup>19)</sup> and an aryl bromide **7**<sup>20)</sup> was carried out as a model system. Although a small amount of **5a** was detected in the presence of excess copper dust<sup>21)</sup> and *N,N*-dimethylformamide (DMF), a serious amount of unavoidable byproducts, such as a dehalogenated compound of **7** and a homocoupling product of **7** were produced while the starting material **6a** remained. This is a typical drawback of the classical Ullmann reaction. Thus, we examined another reagent system using copper(I) species.<sup>22)</sup> However, the desired **5a** was not obtained and debenylation of the substrates was observed. This complexity might be due to the steric hindrance of the *ortho*-substituted **7**. In this regard, we thought that there would be a possibility only using a less sterically-hindered compound as the coupling partner under the Cu(0)-mediated classical Ullmann reaction to construct the sterically-hindered biaryl ether.

In order to consume phenol **6a** and complete the reaction, we needed 3 eq of the aryl bromide **7** with an excess amount of copper dust in a refluxing solvent.<sup>23)</sup> When we used *N,N*-dimethylacetamide (DMA) as the solvent, **5a** was successfully obtained in an acceptable yield (Chart 2). In this case, the dehalogenated byproduct produced from **7** was easily separated by standard silica gel column chromatography and **5a** was isolated in an almost pure form. In order to improve the yield, we attempted to modify the protective group on the catechol

Chart 3. Synthesis of Dehydrodigallic Acid (**3**)

part of **6**. When **6b**<sup>24)</sup> was subjected to the reaction, **5b** was obtained in a slightly better yield. It was suggested that the steric hindrance of the phenolic side was related to the reactivity.<sup>2)</sup> We postulated that an acetal type protective group, which can be easily attached to the phenolic hydroxyl groups, would be effective to decrease the steric hindrance. From this standpoint, phenols **6c–e** were prepared<sup>25–27)</sup> and underwent the same reaction. It was found that benzylidene acetal **6d** seemed to be the most suitable for this reaction than the orthoester **6c** and *p*-methoxybenzylidene acetal **6e**. We thought that the lower yields of both **5c** and **5e** were due to the instability of the acetal moiety under the reaction conditions. Consequently, the biaryl ether **5d**, which is an equivalent of **5a**, was obtained in a satisfactory yield (91%), and thus, the reaction was carried out on a gram-scale basis.

With the biaryl ether **5d** in hand, we attempted the synthesis of **3** (Chart 3). The transformation of **5d** to Feldman's intermediate **5a** was performed by a simple operation including hydrogenolysis and benzylation. Additionally, the synthesis

Chart 4. Functionalization of Gallic Acid (**1**)Chart 5. Synthesis of **15**

of **3** was also accomplished from **5d** by a two-step sequence that involved the hydrolysis of the methyl ester and reductive cleavage of the benzyl and benzylidene groups. The spectral data of the synthetic **3** was matched the reported data.<sup>28)</sup>

When we plan to synthesize the natural ellagitannin, coriariin B, the two carboxy groups of **3** should be distinguished for the construction of the dehydrodigalloyl part. In order to mask the carboxylic acid, we selected the benzyl and *tert*-butyl esters, which can be cleaved in a different deprotective manner. Chart 4 shows the preparation of the bromide **9** and phenol **11**. Benzylation of an inexpensive starting material **1** (monohydrate) afforded **8**,<sup>29)</sup> which was smoothly brominated to give **9**. On the other hand, the *tert*-butylation of **1** was accomplished using Ohta's system.<sup>30)</sup> It was noted that although the yield of **10**<sup>31)</sup> was quite low, this method was the best way for our intensive examination because the production of any undesired byproducts was suppressed and the unreacted **1** was easily removed. For the protection of the catechol moiety, the reaction of **10** with benzaldehyde in pyridine was carried out to provide **11**.

The phenol **11** and bromide **9** were reacted by heating at 160°C to afford the biaryl ether **12**, which was successively deprotected without rigorous purification (Chart 5). Selective removal of the benzylidene group was achieved by use of the SnCl<sub>2</sub>-EtSH combination system<sup>32)</sup> to give **13** in 76% yield. Compound **14** could be obtained by the reprotection of the catechol part of **13**. Finally, the *tert*-butyl group of **14** was removed by SnCl<sub>2</sub>, which afforded the desired carboxylic acid **15**. Thus, we succeeded in constructing the synthetic compo-

nent of coriariin B.

In conclusion, we investigated the formation of a highly functionalized biaryl ether derived from **1** via the classical Ullmann reaction. By using the benzylidene group, the biaryl ether **5d** was sufficiently obtained, and dehydrodigallic acid (**3**) was readily synthesized. Additionally, we demonstrated the construction of the DHDG part of coriariin B. We believe that compound **15**, which is derivative of **3**, must be a common building block toward the synthesis of the ellagitannins.

## Experimental

Melting points were measured using a Yanagimoto micro melting point hot-plate and are uncorrected. The IR spectra were recorded using a SHIMADZU FTIR-8400 spectrophotometer. The NMR spectra were obtained using a JEOL  $\alpha$ -400 (400 MHz) or a JEOL JNX-ECX500 (500 MHz) instrument. The chemical shifts are given in  $\delta$  ppm with tetramethylsilane (TMS) as an internal standard. The elemental analyses were performed using a Thermo Scientific FlashEA1112 analyzer. FAB-MS was obtained using a JEOL JMS-AX505HAD instrument with *m*-nitrobenzyl alcohol as the matrix. Electron ionization-mass spectra (EI-MS) was obtained using a JMS-GCmate II instrument. Silica gel column chromatography was carried out using a wakogel® C-200 (Wako). TLC analysis was performed on Kieselgel 60 F<sub>254</sub> (Merck) plates. Copper dust was activated by the reported method.<sup>21)</sup> Solvents were dried by the standard procedure.

**General Procedure for Ullmann Reaction** A mixture of ArOH (**6a–e**) (1.00 mmol), ArBr (**7**) (1.60 g, 3.00 mmol),

and activated Cu (381 mg, 6.00 mmol) in DMA (2.5 mL) was refluxed under N<sub>2</sub> in a pre-heated oil bath at 200°C. After stirring for 15–30 min, the reaction mixture was cooled to ambient temperature, diluted with EtOAc, and filtered. The filtrate was poured into H<sub>2</sub>O and extracted with EtOAc. The combined organic layer was washed with brine, dried over MgSO<sub>4</sub>, and filtered. The filtrate was evaporated and the resulting residue was purified by silica gel column chromatography (1:4–1:3; EtOAc–hexane), providing the biaryl ether product (**5a–e**).

Methyl 3,4,5-Tribenzyloxy-2-(2,3-dibenzyloxy-5-methoxycarbonylphenoxy)benzoate<sup>10</sup> (**5a**): From **6a** (455 mg, 1.00 mmol), **5a** (547 mg, *ca.* 67%, yellow oil) was obtained by the same procedure mentioned above. The reaction was carried out for 30 min; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>) δ: 3.72 (3H, s, OMe), 3.80 (3H, s, OMe), 4.97 (2H, s, CH<sub>2</sub>), 5.13 (4H, s, CH<sub>2</sub>), 5.14 (2H, s, CH<sub>2</sub>), 5.15 (2H, s, CH<sub>2</sub>), 6.91 (1H, d, *J*=1.5 Hz, ArH), 7.11–7.47 (27H, m, ArH); IR (CHCl<sub>3</sub>) ν<sub>max</sub> 3033, 3015, 2953, 2357, 1717, 1539, 1435, 1338, 1224, 1206, 1093, 1011, 770, 697, 666 cm<sup>-1</sup>.

Methyl 3,4,5-Tribenzyloxy-2-(2,3-dimethoxy-5-methoxycarbonylphenoxy)benzoate (**5b**): From **6b** (212 mg, 1.00 mmol), **5b** (527 mg, 79%, pale yellow solid) was obtained by the same procedure mentioned above. The reaction was carried out for 30 min; mp 142–144°C; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>) δ: 3.73 (3H, s, OMe), 3.81 (3H, s, OMe), 3.94 (3H, s, OMe), 3.94 (3H, s, OMe), 4.96 (2H, s, CH<sub>2</sub>), 5.13 (2H, s, CH<sub>2</sub>), 5.14 (2H, s, CH<sub>2</sub>), 6.84 (1H, d, *J*=1.5 Hz, ArH), 7.10–7.48 (17H, m, ArH). <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>) δ: 52.3, 52.4, 56.4, 61.1, 71.5, 75.7, 75.8, 107.3, 108.8, 111.0, 119.9, 125.0, 127.9, 128.1, 128.3, 128.3, 128.4, 128.4, 128.5, 128.8, 128.8, 136.4, 136.9, 136.9, 142.0, 142.6, 146.7, 146.9, 150.1, 152.4, 153.4, 165.3, 166.6; IR (KBr) ν<sub>max</sub> 2948, 2360, 2334, 1721, 1595, 1467, 1437, 1417, 1337, 1224, 1097, 1012, 748, 697, 668, 420 cm<sup>-1</sup>; high resolution (HR)-MS (EI) Calcd for C<sub>39</sub>H<sub>36</sub>O<sub>10</sub> [M]<sup>+</sup>: 664.2308; Found: 664.2277 [M]<sup>+</sup>. Anal. Calcd for C<sub>39</sub>H<sub>36</sub>O<sub>10</sub>: C, 70.47; H, 5.46. Found: C, 70.74; H, 5.41.

Methyl 3,4,5-Tribenzyloxy-2-(2,3-ethoxymethylenedioxy-5-methoxycarbonylphenoxy)benzoate (**5c**): From **6c** (240 mg, 1.00 mmol), **5c** (494 mg, 71%, colorless oil) was obtained by the same procedure mentioned above. The reaction was carried out for 15 min; <sup>1</sup>H-NMR (500 MHz, acetone-*d*<sub>6</sub>) δ: 1.19 (3H, t, *J*=6.5 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 3.72 (2H, q, *J*=6.5 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 3.73 (3H, s, OMe), 3.78 (3H, s, OMe), 5.01 (1H, d, A of AB, *J*=11.0 Hz, CH<sub>A</sub>H<sub>B</sub>), 5.03 (1H, d, B of AB, *J*=11.0 Hz, CH<sub>A</sub>H<sub>B</sub>), 5.21 (2H, s, CH<sub>2</sub>), 5.25 (2H, s, CH<sub>2</sub>), 7.03 (1H, d, *J*=1.5 Hz, ArH), 7.09 (1H, s), 7.48 (1H, s), 7.20–7.59 (16H, m, ArH); <sup>13</sup>C-NMR (125 MHz, acetone-*d*<sub>6</sub>) δ: 15.1, 52.4, 52.4, 60.3, 71.8, 76.0, 76.2, 103.9, 111.3, 112.5, 120.8, 121.3, 124.9, 128.7, 128.8, 128.8, 128.9, 129.0, 129.1, 129.3, 129.4, 137.6, 137.9, 138.0, 138.6, 142.7, 142.8, 147.3, 147.3, 148.5, 151.1, 165.3, 166.2; IR (CHCl<sub>3</sub>) ν<sub>max</sub> 3030, 3015, 2953, 1714, 1635, 1438, 1372, 1313, 1212, 1085, 769, 697, 666 cm<sup>-1</sup>; HR-MS (EI) Calcd for C<sub>40</sub>H<sub>36</sub>O<sub>11</sub> [M]<sup>+</sup>: 692.2258; Found: 692.2225 [M]<sup>+</sup>.

Methyl 2-(2,3-Benzaloxo-5-methoxycarbonylphenoxy)-3,4,5-tribenzyloxybenzoate (**5d**): The mixture of **6d** (2.72 g, 10.0 mmol), **7** (16.0 g, 30.0 mmol), and activated Cu (3.81 g, 60.0 mmol) in DMA (25 mL) was refluxed under N<sub>2</sub> in a pre-heated oil bath at 200°C. After stirring for 15 min, the reaction mixture was cooled to ambient temperature, diluted

with EtOAc (400 mL), and filtered using celite. The filtrate was poured into H<sub>2</sub>O (400 mL) and extracted with EtOAc (400 mL×3). The combined organic layer was washed with brine (300 mL), dried over MgSO<sub>4</sub>, and filtered. The filtrate was evaporated and the resulting residue (17.6 g) was purified by silica gel column chromatography (1:4; EtOAc–hexane), providing **5d** (6.60 g, 91%) as a yellow oil; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>) δ: 3.74 (3H, s, OMe), 3.81 (3H, s, OMe), 4.96 (2H, s, CH<sub>2</sub>), 5.09 (2H, s, CH<sub>2</sub>), 5.12 (2H, s, CH<sub>2</sub>), 6.95 (1H, s, PhCH), 7.03 (1H, d, *J*=2.0 Hz, ArH), 7.24 (1H, d, *J*=2.0 Hz, ArH), 7.13–7.53 (21H, m, ArH); <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>) δ: 52.2, 52.4, 71.5, 75.7, 75.8, 104.3, 110.9, 112.0, 112.5, 119.9, 124.3, 126.5, 127.9, 128.0, 128.2, 128.3, 128.4, 128.4, 128.7, 128.8, 130.5, 135.6, 136.4, 136.9, 137.0, 139.7, 142.1, 142.8, 146.7, 146.8, 149.3, 150.2, 165.3, 166.3; IR (CHCl<sub>3</sub>) ν<sub>max</sub> 3030, 3011, 2953, 1712, 1635, 1499, 1437, 1363, 1226, 1201, 1087, 1018, 909, 747, 697 cm<sup>-1</sup>; HR-MS (EI) Calcd for C<sub>44</sub>H<sub>36</sub>O<sub>10</sub> [M]<sup>+</sup>: 724.2308; Found: 724.2283 [M]<sup>+</sup>.

Methyl 3,4,5-Tribenzyloxy-2-(2,3-*p*-methoxybenzaloxo-5-methoxycarbonylphenoxy)benzoate (**5e**): From **6e** (302 mg, 1.00 mmol), **5e** (346 mg, 46%, yellow amorphous) was obtained by the same procedure mentioned above. The reaction was carried out for 15 min; <sup>1</sup>H-NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ: 3.70 (3H, s, OMe), 3.75 (3H, s, OMe), 3.77 (3H, s, OMe), 4.85 (1H, d, A of AB, *J*=10.8 Hz, CH<sub>A</sub>H<sub>B</sub>), 4.92 (1H, d, B of AB, *J*=10.8 Hz, CH<sub>A</sub>H<sub>B</sub>), 5.09 (2H, s, CH<sub>2</sub>), 5.21 (2H, s, CH<sub>2</sub>), 6.82 (1H, s, PhCH), 6.98–7.51 (22H, m, ArH); <sup>13</sup>C-NMR (125 MHz, acetone-*d*<sub>6</sub>) δ: 52.3, 52.4, 55.6, 71.4, 75.9, 76.1, 104.1, 111.2, 112.8, 112.9, 114.8, 120.7, 124.8, 128.5, 128.7, 128.7, 128.7, 128.8, 128.9, 129.0, 129.0, 129.3, 137.5, 137.9, 137.9, 140.4, 142.9, 142.9, 147.2, 147.2, 150.3, 150.9, 162.3, 165.3, 166.2; IR (CHCl<sub>3</sub>) ν<sub>max</sub> 3031, 3013, 2953, 2357, 1714, 1635, 1499, 1436, 1306, 1252, 1223, 1214, 1172, 1086, 1024, 773, 668 cm<sup>-1</sup>; HR-MS (EI) Calcd for C<sub>45</sub>H<sub>38</sub>O<sub>11</sub> [M]<sup>+</sup>: 754.2415; Found: 754.2378 [M]<sup>+</sup>.

**Conversion of 5d to 5a** To a solution of **5d** (643 mg, 0.887 mmol) in MeOH (8 mL) and EtOAc (8 mL), 10% Pd–C (64 mg) was added at room temperature. After stirring for 30 min under H<sub>2</sub>, the reaction mixture was filtered and concentrated *in vacuo* to give a yellow amorphous (371 mg) powder, which was used in the next reaction without further purification. A mixture of the above residue (371 mg) and K<sub>2</sub>CO<sub>3</sub> (1.23 g, 8.90 mmol) in DMF (20 mL) was stirred at room temperature. BnBr (1.06 mL, 8.92 mol) was added to the solution and the mixture was heated at 80°C with stirring. After 1 h, the reaction mixture was poured into H<sub>2</sub>O (100 mL) and extracted with EtOAc (50 mL×3). The combined organic layer was washed with brine (50 mL), dried over MgSO<sub>4</sub>, and filtered. The filtrate was evaporated and the resulting residue (1.63 g) was purified by silica gel column chromatography (1:4; EtOAc–hexane), providing **5a** (661 mg, 91%) as a pale yellow solid.

**Dehydrodigallic Acid (3)** To a solution of **5d** (1.45 g, 2.00 mmol) in THF (10 mL), 10% KOH aq. (10 mL) was added and the mixture was stirred at room temperature. After 18 h, MeOH (5 mL) was added and the mixture was heated at 60°C with stirring for 24 h. The reaction mixture was quenched with 10% HCl aq. (10 mL) and extracted with EtOAc (50 mL×3). The combined organic layer was washed with brine (50 mL), dried over MgSO<sub>4</sub>, and filtered. The filtrate was evaporated and the obtained pale yellow solid (1.45 g)

was recrystallized from EtOAc–hexane, providing the dicarboxylic acid (1.29 g, 93%) as a colorless solid; mp 215–217°C (EtOAc–hexane);  $^1\text{H-NMR}$  (500 MHz, acetone- $d_6$ )  $\delta$ : 4.99 (1H, d, A of AB,  $J=11.0\text{ Hz}$ ,  $\text{CH}_A\text{H}_B$ ), 5.03 (1H, d, B of AB,  $J=11.0\text{ Hz}$ ,  $\text{CH}_A\text{H}_B$ ), 5.18 (2H, s,  $\text{CH}_2$ ), 5.26 (2H, s,  $\text{CH}_2$ ), 7.08 (1H, d,  $J=1.0\text{ Hz}$ , ArH), 7.13 (1H, s, PhCH), 7.19–7.61 (22H, m, ArH);  $^{13}\text{C-NMR}$  (125 MHz, acetone- $d_6$ )  $\delta$ : 71.8, 75.9, 76.1, 104.3, 111.6, 112.7, 113.1, 121.2, 125.3, 127.3, 128.7, 128.8, 128.8, 128.9, 129.0, 129.1, 129.3, 129.4, 129.5, 131.3, 136.7, 137.7, 138.0, 138.0, 140.4, 143.1, 147.2, 147.3, 150.1, 151.0, 165.7, 166.7; IR (KBr)  $\nu_{\text{max}}$  3033, 2945, 2362, 1685, 1631, 1498, 1438, 1367, 1305, 1218, 1089, 1016, 737, 695  $\text{cm}^{-1}$ ; HR-MS (EI) Calcd for  $\text{C}_{42}\text{H}_{32}\text{O}_{10}$   $[\text{M}]^+$ : 696.1995; Found: 696.2034  $[\text{M}]^+$ . The resulting product (139 mg, 0.200 mmol) was dissolved in MeOH (6 mL), 10% Pd–C (14.0 mg) was added to the solution and the mixture was stirred at room temperature under  $\text{H}_2$ . After 2 h, the reaction mixture was filtered and concentrated *in vacuo*, providing **3** (66.7 mg, 99%) as a brown solid;  $^1\text{H-NMR}$  (500 MHz, DMSO- $d_6$ )  $\delta$ : 6.46 (1H, d,  $J=1.5\text{ Hz}$ , ArH), 6.90 (1H, s, ArH), 7.00 (1H, d,  $J=1.5\text{ Hz}$ , ArH);  $^{13}\text{C-NMR}$  (125 MHz, DMSO- $d_6$ )  $\delta$ : 106.6, 108.2, 110.5, 115.2, 119.9, 136.0, 139.0, 139.2, 139.7, 142.5, 145.6, 147.5, 166.3, 167.5; IR (KBr)  $\nu_{\text{max}}$  3498, 2357, 2342, 1685, 1611, 1522, 1440, 1329, 1244, 1190, 1101, 1034, 668  $\text{cm}^{-1}$ ; HR-MS (EI) Calcd for  $\text{C}_{14}\text{H}_{10}\text{O}_{10}$   $[\text{M}]^+$ : 338.0274; Found: 338.0289  $[\text{M}]^+$ . The spectral data of the synthetic **3** matched the data provided in the literature.<sup>26)</sup>

**Benzyl 3,4,5-Tribenzyloxybenzoate**<sup>29)</sup> (**8**) A mixture of gallic acid monohydrate (**1**) (100 g, 0.532 mol),  $\text{K}_2\text{CO}_3$  (587 g, 4.25 mol), and  $^n\text{Bu}_4\text{NI}$  (9.9 g, 26.8 mmol) in DMF (1 L) was stirred using a mechanical stirrer at room temperature. BnCl (501 mL, 4.25 mol) was added to the suspension and the mixture was heated at 100°C with stirring for 4.5 h. The reaction mixture was cooled to ambient temperature, poured into the mixed solution of  $\text{H}_2\text{O}$  (5 L) and hexane (0.5 L), and stirred. The precipitate was then collected on a glass filter and washed with  $\text{H}_2\text{O}$  and hexane. The obtained white solid (497.2 g) was recrystallized from acetone, providing **8** (243 g, 86%) as colorless needles; mp 101.3–102.8°C (acetone);  $^1\text{H-NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$ : 5.10 (4H, s,  $\text{CH}_2$ ), 5.15 (2H, s,  $\text{CH}_2$ ), 5.33 (2H, s,  $\text{CH}_2$ ), 7.26–7.44 (22H, m, ArH);  $^{13}\text{C-NMR}$  (125 MHz,  $\text{CDCl}_3$ )  $\delta$ : 66.8, 71.2, 75.1, 109.2, 125.2, 127.6, 128.0, 128.0, 128.1, 128.2, 128.2, 128.6, 128.6, 136.1, 136.7, 137.5, 142.5, 152.6, 165.9; IR (KBr)  $\nu_{\text{max}}$  3064, 3032, 2945, 2866, 1710, 1594, 1500, 1454, 1428, 1385, 1366, 1338, 1204, 1128, 1029, 971, 862, 753, 730, 695, 604  $\text{cm}^{-1}$ ; HR-MS (EI) Calcd for  $\text{C}_{35}\text{H}_{30}\text{O}_5$   $[\text{M}]^+$ : 530.2093; Found: 530.2115  $[\text{M}]^+$ .

**Benzyl 3,4,5-Tribenzyloxy-2-bromobenzoate** (**9**) To a stirring solution of **8** (79.6 g, 150 mmol) in DMF (350 mL), a solution of *N*-bromosuccinimide (NBS) (30.7 g, 172.5 mmol) in DMF (70 mL) was dropwise added at room temperature. After stirring for 24 h,  $\text{H}_2\text{O}$  was added to the solution and the resulting mixture was then stirred. The precipitate was collected on a glass filter and washed with  $\text{H}_2\text{O}$  and hexane. The obtained solid (112.4 g) was recrystallized from acetone, providing **9** (78.5 g, 86%) as colorless needles; mp 123.8–124.4°C (acetone);  $^1\text{H-NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$ : 5.03 (2H, s,  $\text{CH}_2$ ), 5.10 (4H, s,  $\text{CH}_2$ ), 5.37 (2H, s,  $\text{CH}_2$ ), 7.29–7.51 (21H, m, ArH);  $^{13}\text{C-NMR}$  (125 MHz,  $\text{CDCl}_3$ )  $\delta$ : 67.6, 71.4, 75.6, 75.9, 110.6, 112.4, 127.7, 128.4, 128.4, 128.5, 128.5, 128.6, 128.6, 128.8, 128.8, 128.8, 135.6, 136.1, 136.7, 136.9, 146.0, 151.1, 151.8,

165.9; IR (KBr)  $\nu_{\text{max}}$  3030, 2362, 2333, 1700, 1577, 1454, 1369, 1329, 1216, 1159, 1100, 955, 909, 740, 698  $\text{cm}^{-1}$ ; HR-MS (EI) Calcd for  $\text{C}_{35}\text{H}_{29}\text{O}_5\text{Br}$   $[\text{M}]^+$ : 608.1198; Found: 608.1241  $[\text{M}]^+$ . *Anal.* Calcd for  $\text{C}_{35}\text{H}_{29}\text{O}_5\text{Br}$ : C, 68.97; H, 4.80. Found: C, 68.74; H, 4.75.

**tert-Butyl Gallate**<sup>31)</sup> (**10**) A mixture of gallic acid monohydrate (**1**) (47.0 g, 250 mmol), *t*-BuOH (300 mL), and *t*-BuOAc (300 mL) was stirred at 0°C under  $\text{N}_2$ . Conc.  $\text{H}_2\text{SO}_4$  (25 mL) was carefully added to the solution with maintaining the temperature at 0°C. After stirring for 1 h at the same temperature, the resulting mixture was warmed to room temperature and stirred for 5 d. The reaction mixture was quenched with sat.  $\text{NaHCO}_3$  aq., poured into  $\text{H}_2\text{O}$  (500 mL), and extracted with  $\text{Et}_2\text{O}$  (500 mL  $\times$  3). The combined organic layer was washed with brine (300 mL), dried over  $\text{MgSO}_4$ , and filtered. The filtrate was evaporated and the resulting yellow residue (64.5 g) was dissolved in  $\text{CHCl}_3$  (100 mL). After removal of the insoluble precipitate by filtration, the filtrate was concentrated *in vacuo* and the yellow residue (29.5 g) was separated by silica gel column chromatography (1:4; EtOAc–hexane). The obtained pale yellow solid (24.5 g) was recrystallized from  $\text{CHCl}_3$ , providing **10** (21.7 g, 38%) as a colorless solid. Analytical sample of **10** was obtained by further recrystallization from  $\text{CHCl}_3$ –hexane; mp 146–147°C ( $\text{CHCl}_3$ –hexane); IR (KBr)  $\nu_{\text{max}}$  3377, 3314, 3227, 2980, 1697, 1670, 1624, 1445, 1371, 1348, 1256, 1157, 1026, 771  $\text{cm}^{-1}$ ;  $^1\text{H-NMR}$  (500 MHz, acetone- $d_6$ )  $\delta$ : 1.53 (9H, s, *Ot*-Bu), 7.07 (2H, s, ArH);  $^{13}\text{C-NMR}$  (125 MHz, acetone- $d_6$ )  $\delta$ : 28.3, 80.3, 109.7, 123.6, 138.2, 145.8, 166.0; *Anal.* Calcd for  $\text{C}_{11}\text{H}_{14}\text{O}_5$ : C, 58.40; H, 6.24. Found: C, 58.35; H, 6.40.

**tert-Butyl 3,4-Benzaloxo-5-hydroxybenzoate** (**11**) To a solution of **10** (10.0 g, 44.2 mmol) in pyridine (25.0 mL), benzal chloride (8.54 mL, 66.3 mmol) was added and the mixture was heated at 110°C under  $\text{N}_2$ . After stirring for 8 h, toluene was added to the reaction mixture and the solvent was removed *in vacuo*. The resulting purple residue was dissolved in  $\text{CHCl}_3$  and filtered with  $\text{SiO}_2$  pad. After concentration of the filtrate, pale yellow solid (16.2 g) was recrystallized from  $\text{CHCl}_3$ –hexane, providing **11** (9.71 g, 70%) as a colorless solid; mp 167.5–169.8°C ( $\text{CHCl}_3$ –hexane); IR (KBr)  $\nu_{\text{max}}$  3369, 2976, 1684, 1645, 1614, 1512, 1447, 1398, 1337, 1335, 1305, 1259, 1219, 1163, 1076, 1076, 1016, 766, 696  $\text{cm}^{-1}$ ;  $^1\text{H-NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 1.57 (9H, s, *Ot*-Bu), 7.04 (1H, s, PhCH), 7.13 (1H, d,  $J=1.6\text{ Hz}$ , ArH), 7.33 (1H, d,  $J=1.6\text{ Hz}$ , ArH), 7.44–7.57 (5H, m, ArH);  $^{13}\text{C-NMR}$  (125 MHz, acetone- $d_6$ )  $\delta$ : 28.2, 81.0, 102.4, 111.9, 114.5, 127.1, 127.2, 129.5, 131.2, 137.0, 139.0, 140.9, 149.7, 165.3; *Anal.* Calcd for  $\text{C}_{18}\text{H}_{18}\text{O}_5$ : C, 68.78; H, 5.77. Found: C, 68.50; H, 5.86.

**Benzyl 2-(2,3-Benzaloxo-5-*tert*-butoxycarbonylphenoxy)-3,4,5-tribenzyloxybenzoate** (**12**) Two round-bottomed flasks (200 mL) were equipped with **11** (3.15 g, 10.0 mmol), **9** (18.3 g, 30.0 mmol), activated Cu (3.81 g, 60.0 mmol), and DMA (25 mL), respectively. The each mixture was heated at 160°C under  $\text{N}_2$ . After stirring for 30 min, the each reaction mixture was cooled to ambient temperature, diluted with EtOAc (250 mL), and filtered. The filtrate were combined, poured into  $\text{H}_2\text{O}$  (500 mL), and extracted with EtOAc (500 mL  $\times$  3). The combined organic layer was washed with brine (300 mL), dried over  $\text{MgSO}_4$ , and filtered. The filtrate was evaporated and the resulting residue (46.3 g) was purified by silica gel column chromatography (1:4; EtOAc–hexane) to

give a colorless residue (39.9 g) as a mixture containing **12**, which was used in the next reaction without further purification. Analytical sample of **12** was separately obtained by silica gel column chromatography as a colorless oil;  $^1\text{H-NMR}$  (500 MHz, acetone- $d_6$ )  $\delta$ : 1.52 (9H, s, *Or*-Bu), 4.98 (1H, d, A of AB,  $J=10.5$  Hz,  $\text{CH}_A\text{H}_B$ ), 5.01 (1H, d, B of AB,  $J=10.5$  Hz,  $\text{CH}_A\text{H}_B$ ), 5.17 (2H, s,  $\text{CH}_2$ ), 5.19 (1H, d, A of AB,  $J=12.5$  Hz,  $\text{CH}_A\text{H}_B$ ), 5.24 (1H, d, B of AB,  $J=12.5$  Hz,  $\text{CH}_A\text{H}_B$ ), 5.25 (2H, s,  $\text{CH}_2$ ), 6.98 (1H, d,  $J=1.5$  Hz, ArH), 7.04 (1H, s, PhCH), 7.10 (1H, d,  $J=1.5$  Hz, ArH), 7.50 (1H, s, ArH), 7.19–7.56 (25H, m, ArH);  $^{13}\text{C-NMR}$  (125 MHz, acetone- $d_6$ )  $\delta$ : 28.2, 67.4, 71.8, 76.0, 76.2, 81.2, 104.0, 111.5, 112.5, 112.7, 120.9, 126.9, 127.2, 128.6, 128.8, 128.9, 129.0, 129.1, 129.3, 129.5, 131.2, 136.7, 136.8, 137.5, 137.8, 137.9, 139.7, 142.8, 142.8, 147.3, 147.4, 150.0, 151.0, 164.9, 165.0; IR (CHCl<sub>3</sub>)  $\nu_{\text{max}}$  3034, 3013, 2490, 1704, 1635, 1498, 1436, 1367, 1217, 1159, 1087, 1017, 770, 697  $\text{cm}^{-1}$ ; HR-MS (FAB, positive ion mode) Calcd for C<sub>53</sub>H<sub>47</sub>O<sub>10</sub> [M+H]<sup>+</sup>: 843.3169; Found: 843.3148 [M+H]<sup>+</sup>.

**Benzyl 3,4,5-Tribenzyloxy-2-(5-*tert*-butoxycarbonyl-2,3-dihydroxyphenoxy)benzoate (13)** To a stirring solution of the above residue (39.9 g) in CH<sub>2</sub>Cl<sub>2</sub> (500 mL), EtSH (14.8 mL, 200 mmol) and SnCl<sub>2</sub> (9.13 g, 50.0 mmol) were added at 0°C. After stirring for 22 h at room temperature, the reaction mixture was filtered and concentrated *in vacuo*. The resulting residue (52.7 g) was purified by silica gel column chromatography (1:3–1:2; EtOAc–hexane), providing **13** (11.4 g, 76%) as a colorless amorphous powder;  $^1\text{H-NMR}$  (500 MHz, acetone- $d_6$ )  $\delta$ : 1.46 (9H, s, *Or*-Bu), 4.99 (2H, s,  $\text{CH}_2$ ), 5.20 (2H, s,  $\text{CH}_2$ ), 5.21 (2H, s,  $\text{CH}_2$ ), 5.25 (2H, s,  $\text{CH}_2$ ), 6.75 (1H, d,  $J=1.5$  Hz, ArH), 7.18 (1H, d,  $J=1.5$  Hz, ArH), 7.49 (1H, s, ArH), 7.23–7.56 (20H, m, ArH);  $^{13}\text{C-NMR}$  (125 MHz, acetone- $d_6$ )  $\delta$ : 28.2, 67.3, 71.8, 76.0, 76.2, 80.5, 107.9, 111.6, 111.8, 121.0, 123.2, 128.6, 128.7, 128.9, 128.9, 129.1, 129.1, 129.1, 129.3, 129.3, 136.9, 137.6, 137.9, 138.0, 139.3, 143.4, 146.3, 147.5, 147.6, 147.9, 150.8, 165.1, 165.6; IR (CHCl<sub>3</sub>)  $\nu_{\text{max}}$  3551, 3034, 3014, 1703, 1615, 1455, 1436, 1367, 1339, 1311, 1207, 1178, 11186, 1068, 966, 788, 697  $\text{cm}^{-1}$ ; HR-MS (EI) Calcd for C<sub>46</sub>H<sub>42</sub>O<sub>10</sub> [M]<sup>+</sup>: 754.2778; Found: 754.2744 [M]<sup>+</sup>.

**Benzyl 3,4,5-Tribenzyloxy-2-(2,3-dibenzyloxy-5-*tert*-butoxycarbonylphenoxy)benzoate (14)** To a stirring solution of **13** (1.51 g, 2.00 mmol) and K<sub>2</sub>CO<sub>3</sub> (691 mg, 5.00 mmol) in DMF (10 mL), BnBr (0.52 mL, 4.38 mmol) was added and the mixture was heated at 40°C. After 2 h, the reaction mixture was cooled to ambient temperature, poured into H<sub>2</sub>O (100 mL), and extracted with EtOAc (50 mL × 3). The combined organic layer was washed with brine (50 mL), dried over MgSO<sub>4</sub>, and filtered. The filtrate was evaporated and the resulting yellow residue (2.20 g) was purified by silica gel column chromatography (1:4; EtOAc–hexane), providing **14** (1.71 g, 91%) as a colorless solid. Analytical sample of **14** was obtained by recrystallization from acetone; mp 129–131°C (acetone);  $^1\text{H-NMR}$  (500 MHz, acetone- $d_6$ )  $\delta$ : 1.49 (9H, s, *Or*-Bu), 4.97 (2H, s,  $\text{CH}_2$ ), 5.01 (2H, s,  $\text{CH}_2$ ), 5.20 (4H, s,  $\text{CH}_2$ ), 5.22 (2H, s,  $\text{CH}_2$ ), 5.27 (2H, s,  $\text{CH}_2$ ), 6.86 (1H, d,  $J=2.0$  Hz, ArH), 7.17–7.57 (32H, m, ArH);  $^{13}\text{C-NMR}$  (125 MHz, acetone- $d_6$ )  $\delta$ : 28.2, 67.4, 71.7, 71.8, 75.3, 76.0, 76.1, 81.2, 109.5, 109.6, 111.6, 121.0, 127.8, 128.4, 128.5, 128.6, 128.7, 128.7, 128.8, 128.9, 129.0, 129.1, 129.2, 129.3, 136.8, 137.6, 137.8, 138.0, 138.0, 139.0, 142.1, 142.9, 147.2, 147.6, 150.8, 153.5, 165.0, 165.2; IR (KBr)  $\nu_{\text{max}}$  3031, 2952, 2357, 1716, 1594, 1423, 1371, 1343, 1222, 1095, 1000, 967, 859, 753, 697  $\text{cm}^{-1}$ ; HR-MS (FAB, posi-

tive ion mode) Calcd for C<sub>60</sub>H<sub>55</sub>O<sub>10</sub> [M+H]<sup>+</sup>: 935.3795; Found: 935.3816 [M+H]<sup>+</sup>. Anal. Calcd for C<sub>60</sub>H<sub>54</sub>O<sub>10</sub>: C, 77.07; H, 5.82. Found: C, 76.87; H, 5.87.

**3,4-Dibenzyloxy-5-(2,3,4-tribenzyloxy-6-benzyloxycarbonylphenoxy)benzoic Acid (15)** To a stirring solution of **14** (935 mg, 1.00 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL), AcOH (286  $\mu\text{L}$ , 5.00 mmol) and SnCl<sub>2</sub> (457 mg, 2.50 mmol) were subsequently added. After stirring for 22 h at room temperature, AcOH (286  $\mu\text{L}$ , 5.00 mmol) and SnCl<sub>2</sub> (457 mg, 2.50 mmol) were added to the mixture and the resulting mixture was stirred. After 32 h, the reaction mixture was filtered and concentrated *in vacuo*. The obtained yellow amorphous (1.02 g) was purified by silica gel column chromatography (1:1; EtOAc–hexane), providing **15** (777 mg, 88%) as a colorless solid. Analytical sample of **15** was obtained by recrystallization from acetone–hexane; mp 137–139°C (acetone–hexane);  $^1\text{H-NMR}$  (500 MHz, acetone- $d_6$ )  $\delta$ : 4.93 (2H, s,  $\text{CH}_2$ ), 5.01 (2H, s,  $\text{CH}_2$ ), 5.21 (2H, s,  $\text{CH}_2$ ), 5.22 (2H, s,  $\text{CH}_2$ ), 5.25 (2H, s,  $\text{CH}_2$ ), 5.26 (2H, s,  $\text{CH}_2$ ), 6.94 (1H, d,  $J=2.0$  Hz, ArH), 7.16–7.58 (32H, m, ArH);  $^{13}\text{C-NMR}$  (125 MHz, acetone- $d_6$ )  $\delta$ : 67.4, 71.5, 71.6, 75.3, 75.8, 76.0, 109.7, 111.4, 120.8, 126.0, 128.3, 128.5, 128.6, 128.6, 128.8, 128.8, 128.9, 129.0, 129.0, 129.1, 129.2, 129.3, 136.6, 137.3, 137.6, 137.7, 137.7, 138.8, 142.3, 142.7, 147.1, 147.4, 150.7, 153.5, 153.6, 165.0, 167.3; IR (KBr)  $\nu_{\text{max}}$  3064, 3032, 2947, 2874, 2360, 2342, 1704, 1684, 1591, 1497, 1426, 1375, 1217, 1092, 972, 909, 749, 736, 696  $\text{cm}^{-1}$ ; HR-MS (FAB, positive ion mode) Calcd for C<sub>56</sub>H<sub>47</sub>O<sub>10</sub> [M+H]<sup>+</sup>: 879.3169; Found: 879.3190 [M+H]<sup>+</sup>.

**Acknowledgments** This work was financially supported by the JSPS KAKENHI Grant No. 22590003 and the Tamura Science and Technology Foundation.

## References and Notes

- 1) "Chemistry and Biology of Ellagitannins. An Underestimated Class of Bioactive Plant Polyphenols," ed. by Quideau S., World Scientific Publishing Co., Pte. Ltd., Singapore, 2009.
- 2) Quideau S., Feldman K. S., *Chem. Rev.*, **96**, 475–503 (1996).
- 3) Khanbabaee K., van Ree T., *Synthesis*, 1585–1610 (2001).
- 4) Feldman K. S., *Phytochemistry*, **66**, 1984–2000 (2005).
- 5) Pouységu L., Deffieux D., Malik G., Natangelo A., Quideau S., *Nat. Prod. Rep.*, **28**, 853–874 (2011).
- 6) Mayer W., *Liebigs Ann.*, **578**, 33–44 (1952).
- 7) Hatano T., Hattori S., Okuda T., *Chem. Pharm. Bull.*, **34**, 4092–4097 (1986).
- 8) Mayer W., Fikentscher R., *Chem. Ber.*, **91**, 1542–1545 (1958).
- 9) Feldman K. S., Quideau S., Appel H. M., *J. Org. Chem.*, **61**, 6656–6665 (1996).
- 10) Feldman K. S., Sahasrabudhe K., *J. Org. Chem.*, **64**, 209–216 (1999).
- 11) Feldman K. S., Lawlor M. D., *J. Am. Chem. Soc.*, **122**, 7396–7397 (2000).
- 12) Feldman K. S., Lawlor M. D., Sahasrabudhe K., *J. Org. Chem.*, **65**, 8011–8019 (2000).
- 13) Our preliminary attempts on hydrolysis conditions to distinguish the two methyl esters were unsuccessful.
- 14) Ley S. V., Thomas A. W., *Angew. Chem. Int. Ed.*, **42**, 5400–5449 (2003), and references cited therein.
- 15) The basic examination of the Ullmann condensation was performed in our previous studies.<sup>16–18)</sup>
- 16) Abe H., Sahara Y., Matsuzaki Y., Takeuchi Y., Harayama T., *Tetrahedron Lett.*, **49**, 605–609 (2008).
- 17) Shioe K., Takeuchi Y., Harayama T., Abe H., *Chem. Pharm. Bull.*,

- 58, 435–437 (2010).
- 18) Shioe K., Sahara Y., Horino Y., Harayama T., Takeuchi Y., Abe H., *Tetrahedron*, **67**, 1960–1970 (2011).
- 19) Pearson A. J., Bruhn P. R., *J. Org. Chem.*, **56**, 7092–7097 (1991).
- 20) Ohzeki T., Mori K., *Biosci. Biotechnol. Biochem.*, **67**, 2584–2590 (2003).
- 21) Fuson R. C., Cleveland E. A., *Org. Synth., Coll. Vol. III*, 339–340 (1955).
- 22) Maiti D., Buchwald S. L., *J. Org. Chem.*, **75**, 1791–1794 (2010).
- 23) Because the homo-coupling of the aryl bromide could not be avoided, 3 eq. of **7** was necessary.
- 24) Tanaka M., Ikeya Y., Mitsuhashi H., Maruno M., Wakamatsu T., *Tetrahedron*, **51**, 11703–11724 (1995).
- 25) Percec V., Bera T. K., Glodde M., Fu Q., Balagurusamy V. S. K., Heiney P. A., *Chemistry*, **9**, 921–935 (2003).
- 26) Domon L., Uguen D., *Tetrahedron Lett.*, **41**, 5501–5505 (2000).
- 27) Alam A., Takaguchi Y., Ito H., Yoshida T., Tsuboi S., *Tetrahedron*, **61**, 1909–1918 (2005).
- 28) Nawwar M. A. M., Hussein S. A. M., Buddrus J., Linscheid M., *Phytochemistry*, **35**, 1349–1354 (1994).
- 29) Thapa M., Kim Y., Desper J., Chang K., Hua D. H., *Bioorg. Med. Chem. Lett.*, **22**, 353–356 (2012).
- 30) Ohta S., Shimabayashi A., Makino N., Okamoto M., *Yakugaku Zasshi*, **103**, 991–993 (1983).
- 31) Wijesekera R. O. B., Ratnayake V. U., *J. Appl. Chem. Biotechnol.*, **22**, 559–564 (1972).
- 32) Xia J., Hui Y., *Synth. Commun.*, **26**, 881–886 (1996).